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(21) International Application Number: PCT/US97/12410		(72) Inventors; and (75) Inventors/Applicants (for US only): TSIEN, Roger, Y. [US/US]; 8535 Nottingham Place, La Jolla, CA 92037 (US). CUBITT, Andrew, B. [GB/US]; 11083 Caminito Encanto, San Diego, CA 92131 (US).	
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(71) Applicant (for all designated States except US): THE REGENTS OF THE UNIVERSITY OF CALIFORNIA [US/US]; 22nd floor, 300 Lakeside Drive, Oakland, CA 94612-3550 (US).			
(54) Title: ASSAYS FOR PROTEIN KINASES USING FLUORESCENT PROTEIN SUBSTRATES			
(57) Abstract This invention provides assays for protein kinase activity using fluorescent proteins engineered to include sequences that can be phosphorylated by protein kinases. The proteins exhibit different fluorescent properties in the non-phosphorylated and phosphorylated states.			

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ASSAYS FOR PROTEIN KINASES USING FLUORESCENT PROTEIN SUBSTRATES

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BACKGROUND OF THE INVENTION

This invention relates to the field of enzymatic assays and, in particular, assays for protein kinase activity involving modified fluorescent proteins.

Protein phosphorylation is one of the most important general mechanisms of cellular regulation. Protein phosphorylation commonly occurs on three major amino acids, tyrosine, serine or threonine, and changes in the phosphorylation state of these amino acids within proteins can regulate many aspects of cellular metabolism, regulation, growth and differentiation. Changes in the phosphorylation state of proteins, mediated through phosphorylation by kinases, or dephosphorylation by phosphatases, is a common mechanism through which cell surface signaling pathways transmit and integrate information into the nucleus. Given their key role in cellular regulation, it is not surprising that defects in protein kinases and phosphatases have been implicated in many disease states and conditions. For example, the over-expression of cellular tyrosine kinases such as the EGF or PDGF receptors, or the mutation of tyrosine kinases to produce constitutively active forms (oncogenes) occurs in many cancer cells. Drucker et al. (1996) *Nature Medicine* 2: 561-56. Protein tyrosine kinases are also implicated in inflammatory signals. Defective Thr/Ser kinase genes have been demonstrated to be implicated in several diseases such as myotonic dystrophy as well as cancer, and Alzheimer's disease (Sanpei et al. (1995) *Biochem. Biophys. Res. Commun.* 212: 341-6; Sperber et al (1995) *Neurosci. Lett.* 197: 149-153; Grammas et al (1995) *Neurobiology of Aging* 16: 563-569; Govoni et al. (1996) *Ann. N.Y. Acad. Sci.* 777: 332-337).

The involvement of protein kinases and phosphatases in disease states makes them attractive targets for the therapeutic intervention of drugs, and in fact many clinically useful drugs act on protein kinases or phosphatases. Examples include cyclosporin A which is a potent immunosuppressant that binds to cyclophilin. This complex binds to the Ca/calmodulin-dependent protein phosphatase type 2B (calcineurin) inhibiting its activity, and hence the activation of T-cells. (Sigal and Dumont (1992), Schreiber and Crabtree (1992)). Inhibitors of protein kinase C are in clinical trials as

therapeutic agents for the treatment of cancer. (*Clin. Cancer Res.* (1995) 1:113-122) as are inhibitors of cyclin dependent kinase. (*J. Mol. Med.* (1995) 73:(10):509-14.)

The number of known kinases and phosphatases are growing rapidly as the influence of genomic programs to identify the molecular basis for diseases have increased in size and scope. These studies are likely to implicate many more kinase and phosphatase genes in the development and propagation of diseases in the future, thereby making them attractive targets for drug discovery. However current methods of measuring protein phosphorylation have many disadvantages which prevents or limits the ability to rapidly screen using miniaturized automated formats of many thousands of compounds. This is because many current methods rely on the incorporation and measurement of ^{32}P into the protein substrates of interest. In whole cells this necessitates the use of high levels of radioactivity to efficiently label the cellular ATP pool and to ensure that the target protein is efficiently labeled with radioactivity. After incubation with test drugs, the cells must be lysed and the protein of interest purified to determine its relative degree of phosphorylation. This method requires high numbers of cells, long preincubation times, careful manipulation and washing steps (to avoid artifactual phosphorylation or dephosphorylation), as well as a method of purification of the target protein. Furthermore, final radioactive incorporation into target proteins is usually very low, giving the assay poor sensitivity. Alternative assay methods, for example based on phosphorylation-specific antibodies using ELISA-type approaches, involve the difficulty of producing antibodies that distinguish between phosphorylated and non-phosphorylated proteins, and the requirement for cell lysis, multiple incubation and washing stages which are time consuming, complex to automate and potentially susceptible to artifacts.

Kinase assays based on purified enzymes require large amounts of purified kinases, high levels of radioactivity, and methods of purification of the substrate protein away from incorporated ^{32}P -labelled ATP. They also suffer from the disadvantage of lacking the physiological context of the cell, preventing a direct assessment of a drugs toxicity and ability to cross the cells plasma membrane.

Fluorescent molecules are attractive as reporter molecules in many assay systems because of their high sensitivity and ease of quantification. Recently, fluorescent proteins have been the focus of much attention because they can be produced *in vivo* by biological systems, and can be used to trace intracellular events without the need to be introduced into the cell through microinjection or permeabilization. The green

fluorescent protein of *Aequorea victoria* is particularly interesting as a fluorescent indicator protein. A cDNA for the protein has been cloned. (D.C. Prasher et al., "Primary structure of the *Aequorea victoria* green-fluorescent protein," *Gene* (1992) 111:229-33.) Not only can the primary amino acid sequence of the protein be expressed
5 from the cDNA, but the expressed protein can fluoresce. This indicates that the protein can undergo the cyclization and oxidation believed to be necessary for fluorescence. The fluorescence of green fluorescent protein is generated from residues S65-Y66-G67.

Fluorescent proteins have been used as markers of gene expression, tracers of cell lineage and as fusion tags to monitor protein localization within living cells. (M.
10 Chalfie et al., "Green fluorescent protein as a marker for gene expression," *Science* 263:802-805; A.B. Cubitt et al., "Understanding, improving and using green fluorescent proteins," *TIBS* 20, November 1995, pp. 448-455. U.S. patent 5,491,084, M. Chalfie and D. Prasher. Furthermore, mutant versions of green fluorescent protein have been identified that exhibit altered fluorescence characteristics, including altered excitation and
15 emission maxima, as well as excitation and emission spectra of different shapes. (R. Heim et al., "Wavelength mutations and posttranslational autooxidation of green fluorescent protein," *Proc. Natl. Acad. Sci. USA*, (1994) 91:12501-04; R. Heim et al., "Improved green fluorescence," *Nature* (1995) 373:663-665.) These properties add variety and utility to the arsenal of biologically based fluorescent indicators.

20 There is a need for assays of protein phosphorylation that are simple, sensitive, non-invasive, applicable to living cells and tissues and that avoid the use of any radioactivity.

SUMMARY OF THE INVENTION

25 When fluorescent proteins are modified to incorporate a phosphorylation site recognized by a protein kinase, the fluorescent proteins not only can become phosphorylated by the protein kinase, but they also can exhibit different fluorescent characteristics in their un-phosphorylated and phosphorylated forms when irradiated with light having a wavelength within their excitation spectrum. This characteristic makes
30 fluorescent protein substrates particularly useful for assaying protein kinase activity in a sample.

This invention provides methods for determining whether a sample contains protein kinase activity. The methods involve contacting the sample with a

phosphate donor, usually ATP, and a fluorescent protein substrate of the invention; exciting the fluorescent protein substrate with light of an appropriate wavelength; and measuring the amount of a fluorescent property that differs in the un-phosphorylated state and phosphorylated state. An amount that is consistent with the presence of the
5 fluorescent protein substrate in its phosphorylated state indicates the presence of protein kinase activity, and an amount that is consistent with the presence of the protein substrate in its un-phosphorylated state indicates the absence of protein kinase activity.

One embodiment of the above method is for determining the amount of protein kinase activity in a sample. In this method, measuring the amount of a
10 fluorescent property in the sample comprises measuring the amount at two or more time points after contacting the sample with a phosphate donor and a fluorescent protein substrate of the invention, and determining the quantity of change or rate of change of the measured amount. The quantity or rate of change of the measured amount reflects the amount of protein kinase activity in the sample.

15 In another aspect, the invention provides methods for determining whether a cell exhibits protein kinase activity. The methods involve the steps of providing a transfected host cell of the invention that produces a fluorescent protein substrate of the invention; exciting the protein substrate in the cell with light of an appropriate wavelength; and measuring the amount of a fluorescent property that differs in the un-
20 phosphorylated and phosphorylated states. An amount that is consistent with the presence of the protein substrate in its phosphorylated state indicates the presence of protein kinase activity, and an amount that is consistent with the presence of the protein substrate in its un-phosphorylated state indicates the absence of protein kinase activity or the presence of phosphatase activity.

25 In another aspect, the invention provides methods for determining the amount of activity of a protein kinase in one or more cells from an organism. The methods involve providing a transfected host cell comprising a recombinant nucleic acid molecule comprising expression control sequences operatively linked to a nucleic acid sequence coding for the expression of a fluorescent protein substrate of the invention, the
30 cell expressing the fluorescent protein substrate; exciting the protein substrate in the cell with light; and measuring the amount of a fluorescent property that differs in the un-phosphorylated and phosphorylated states at two or more time points after contacting the sample with a phosphate donor and a fluorescent protein substrate, and determining the

quantity or rate of change of the measured amount. The quantity or rate of change of the measured amount reflects the amount of protein kinase activity in the sample.

This invention also provides screening methods for determining whether a compound alters the activity of a protein kinase. The methods involve contacting a sample containing a known amount of protein kinase activity with the compound, a phosphate donor for the protein kinase and a fluorescent protein substrate of the invention; exciting the protein substrate; measuring the amount of protein kinase activity in the sample as a function of the quantity or rate of change of a fluorescent property that differs in the un-phosphorylated and phosphorylated states; and comparing the amount of activity in the sample with a standard activity for the same amount of the protein kinase. A difference between the amount of protein kinase activity in the sample and the standard activity indicates that the compound alters the activity of the protein kinase.

Another aspect of the drug screening methods involve determining whether a compound alters the protein kinase activity in a cell. The methods involve providing first and second transfected host cells exhibiting protein kinase activity and expressing a fluorescent protein substrate of the invention; contacting the first cell with an amount of the compound; contacting the second cell with a different amount of the compound; exciting the protein substrate in the first and second cells; measuring the amount of protein kinase activity as a function of the quantity of change or rate of change of a fluorescent property that differs in the un-phosphorylated and phosphorylated states in the first and second cells; and comparing the amount in the first and second cells. A difference in the amount indicates that the compound alters protein kinase activity in the cell.

This invention also provides fluorescent protein substrates for a protein kinase. Fluorescent protein substrates for a protein kinase comprise a fluorescent protein moiety and a phosphorylation site for a protein kinase. The protein substrate exhibits a different fluorescent property in the phosphorylated state than in the unphosphorylated state. In a preferred embodiment, the fluorescent protein is an *Aequorea*-related fluorescent protein. In another embodiment, the phosphorylation site is located within about 5, 10, 15 or 20 amino acids of a terminus, e.g., the amino-terminus, of the fluorescent protein moiety. In another embodiment, the protein substrate comprises the phosphorylation site more than 20 amino acids from a terminal of the fluorescent protein moiety and within the fluorescent protein moiety. The phosphorylation site can be one

recognized by, for example, protein kinase A, a cGMP-dependent protein kinase, protein kinase C, Ca^{2+} /calmodulin-dependent protein kinase I, Ca^{2+} / calmodulin-dependent protein kinase II or MAP kinase activated protein kinase type 1.

5 This invention also provides nucleic acid molecules coding for the expression of a fluorescent protein substrate for a protein kinase of the invention. In one aspect, the nucleic acid molecule is a recombinant nucleic acid molecule comprising expression control sequences operatively linked to a nucleic acid sequence coding for the expression of a fluorescent protein substrate for a protein kinase of the invention. In another aspect, the invention provides transfected host cells transfected with a
10 recombinant nucleic acid molecule comprising expression control sequences operatively linked to a nucleic acid sequence coding for the expression of a fluorescent protein substrate for a protein kinase of the invention.

In another aspect, this invention provides collections of fluorescent protein candidate substrates comprising at least 10 different members, each member comprising a
15 fluorescent protein moiety and a variable peptide moiety around the terminus of the fluorescent protein moiety.

In another embodiment, the invention provides collections of recombinant nucleic acid molecules comprising at least 10 different recombinant nucleic acid molecule members, each member comprising expression control sequences operatively linked to
20 nucleic acid sequences coding for the expression of a different fluorescent protein candidate substrate of the invention. The invention also provides collections of host cells comprising at least 10 different host cell members, each member comprising the above recombinant nucleic acid molecules.

The collections of cells are useful in determining the specificity of cellular
25 kinases, from either diseased or normal tissues. The screening methods involve providing a collection of transfected host cells of the invention; culturing the collection of host cells under conditions for the expression of the fluorescent protein candidate substrate; and determining for each of a plurality of members from the collection whether the member contains a fluorescent protein candidate substrate that exhibits a
30 fluorescent property different than the fluorescent property exhibited by the non-phosphorylated candidate substrate. The presence of fluorescent protein candidate substrate that exhibits a fluorescent property different than the fluorescent property

exhibited by the candidate substrate indicates that the candidate substrate possesses a peptide moiety that can be phosphorylated by the kinase present in the host cells.

This invention also provides kits comprising a fluorescent protein substrate and a phosphate donor.

5

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a flow chart showing the steps in an assay method for protein kinase activity.

Fig. 2 depicts molecular events in a cell in altering and detecting
10 fluorescent properties of a fluorescent protein substrate for a protein kinase.

Fig. 3 depicts the nucleotide sequence (SEQ ID NO:1) and deduced amino acid sequence (SEQ ID NO:2) of a wild-type *Aequorea* green fluorescent protein.

Fig. 4 provides a list of the positions and amino acid changes made for phosphorylation mutants made more than fifteen amino acids in the primary sequence
15 from the N-terminus, as compared to Fig. 3. Amino acids underlined represent the phosphorylation motif, amino acids in brackets represent wild type sequence at those positions.

Fig. 5 depicts plasmid pRSET containing a region encoding GFP that is fused in frame with nucleotides encoding an N-terminal polyhistidine tag.

20 Figs. 6A-6E show the fluorescence excitation spectra before and after phosphorylation of N-terminal phosphorylation mutants by protein kinase A using standard phosphorylation conditions. 6A: 1MSRRRRSI (SEQ ID NO:31). 6B: 1MRRRRSII (SEQ ID NO:32). 6C: -1MRRRRSIII (SEQ ID NO:33). 6D: -2MRRRRSIIIF (SEQ ID NO:34). 6E: -3MRRRRSIIIF (SEQ ID NO:35). In all
25 cases the spectrum after phosphorylation has higher amplitude than the spectrum before phosphorylation.

Fig. 7 depicts an expression vector having expression control sequences operably linked to sequences coding for the expression of protein kinase A catalytic subunit (PKA cat) upstream from sequences coding for the expression of a fluorescent
30 protein substrate.

Fig. 8 depicts the fluorescence excitation spectrum of 1MRRRRSII (SEQ ID NO:33): S65A, N149K, V163A and I167T before and after phosphorylation by

protein kinase A using standard phosphorylation conditions. The spectrum after phosphorylation has higher amplitude than the spectrum before phosphorylation.

DETAILED DESCRIPTION OF THE INVENTION

I. METHODS FOR ASSAYING SAMPLES FOR PROTEIN KINASES

Protein kinases add a phosphate residue to the phosphorylation site of a protein, generally through the hydrolysis of ATP to ADP. Fluorescent protein substrates for protein kinases are useful in assays to determine the amount of protein kinase activity in a sample without the need for radioactivity. The assays of this invention take advantage of the fact that phosphorylation of the protein substrate results in a change in a fluorescent property of the fluorescent protein. Methods for determining whether a sample has kinase activity involve contacting the sample with a fluorescent protein substrate having a phosphorylation site recognized by the protein kinase to be assayed and with a phosphate donor under selected test conditions. A phosphate donor is a compound containing a phosphate moiety which the kinase is able to use to phosphorylate the protein substrate. ATP (adenosine-5'-triphosphate) is by far the most common phosphate donor. In certain instances, the sample will contain enough of a phosphate donor to make this step unnecessary. Then the fluorescent protein substrate is excited with light in its excitation spectrum. If the protein substrate has been phosphorylated, the substrate will exhibit different fluorescent properties, indicating that the sample contains protein kinase activity. For example, if the phosphorylated form of the protein substrate has higher fluorescence than the un-phosphorylated form, the amount of fluorescence in the sample will increase as a function of the amount of substrate that has been phosphorylated. If the fluorescent property is a change in the wavelength maximum of emission, the change will be detected as a decrease in fluorescence at the wavelength maximum of the un-phosphorylated substrate and an increase in fluorescence at the wavelength maximum of the phosphorylated substrate.

The amount of kinase activity in a sample can be determined by measuring the amount of a fluorescent property in the sample at a first time and a second time after contact between the sample, the fluorescent protein substrate and a phosphate donor, and determining the degree of change or the rate of change in a fluorescent property. For example, if phosphorylation results in an increase in fluorescence at the excitation wavelength maximum, the fluorescence of the substrate at this wavelength can be

determined at two times. The amount of enzyme activity in the sample can be calculated as a function of the difference in the determined amount of the property at the two times. For example, the absolute amount of activity can be calibrated using standards of enzyme activity determined for certain amounts of enzyme after certain amounts of time. The
5 faster or larger the difference in the amount, the more enzyme activity must have been present in the sample. The amount of a fluorescent property can be determined from any spectral or fluorescence lifetime characteristic of the excited substrate, for example, by determining the intensity of the fluorescent signal from the protein substrate or the excited state lifetime of the protein substrate, the ratio of the fluorescences at two
10 different excitation wavelengths, the ratio of the intensities at two different emission wavelengths, or the excited lifetime of the protein substrate.

Fluorescence in a sample is measured using a fluorimeter. In general, excitation radiation from an excitation source having a first wavelength, passes through excitation optics. The excitation optics cause the excitation radiation to excite the
15 sample. In response, fluorescent proteins in the sample emit radiation which has a wavelength that is different from the excitation wavelength. Collection optics then collect the emission from the sample. The device can include a temperature controller to maintain the sample at a specific temperature while it is being scanned. According to one embodiment, a multi-axis translation stage moves a microtiter plate holding a
20 plurality of samples in order to position different wells to be exposed. The multi-axis translation stage, temperature controller, auto-focusing feature, and electronics associated with imaging and data collection can be managed by an appropriately programmed digital computer. The computer also can transform the data collected during the assay into another format for presentation. This process can be miniaturized and automated to
25 enable screening many thousands of compounds.

Methods of performing assays on fluorescent materials are well known in the art and are described in, e.g., Lakowicz, J.R., *Principles of Fluorescence Spectroscopy*, New York: Plenum Press (1983); Herman, B., Resonance energy transfer microscopy, in: *Fluorescence Microscopy of Living Cells in Culture, Part B, Methods in*
30 *Cell Biology*, vol. 30, ed. Taylor, D.L. & Wang, Y.-L., San Diego: Academic Press (1989), pp. 219-243; Turro, N.J., *Modern Molecular Photochemistry*, Menlo Park: Benjamin/Cummings Publishing Co., Inc. (1978), pp. 296-361.

Enzymatic assays also can be performed on isolated living cells *in vivo*, or from samples derived from organisms transfected to express the substrate. Because fluorescent protein substrates can be expressed recombinantly inside a cell, the amount of enzyme activity in the cell or organism of which it is a part can be determined by determining a fluorescent property or changes in a fluorescent property of cells or samples from the organism.

In one embodiment, shown in Fig 2, a cell is transiently or stably transfected with an expression vector 200 encoding a fluorescent protein substrate containing a phosphorylation site for the enzyme to be assayed. This expression vector optionally includes controlling nucleotide sequences such as promotor or enhancing elements. The expression vector expresses the fluorescent protein substrate 210 that contains the phosphorylation site 211 for the kinase to be detected. The enzyme to be assayed may either be intrinsic to the cell or may be introduced by stable transfection or transient co-transfection with another expression vector encoding the enzyme and optionally including controlling nucleotide sequences such as promoter or enhancer elements. The fluorescent protein substrate and the enzyme preferably are located in the same cellular compartment so that they have more opportunity to come into contact.

If the cell possesses a high degree of enzyme activity ($K = \text{"kinase"}$ 220), the fluorescent protein substrate will be phosphorylated 230 (PO_4), usually through the hydrolysis of ATP. If the cell does not possess kinase activity, or possesses very little, the cell contains substantial amounts of un-phosphorylated substrate 240. Upon excitation with light of the appropriate wavelength ($h\nu_1$) the phosphorylated substrate will fluoresce light ($h\nu_2$). Un-phosphorylated substrate exhibits different fluorescent characteristics upon excitation at the same wavelength, and may, for example, not fluoresce at all, or fluoresce weakly. The amount of the fluorescent property is measured generally using the optics 250 and detector 260 of a fluorimeter.

If the cell contains phosphatases that compete with the protein kinases, removing the phosphate from the protein substrate, the level of enzyme activity in the cell can reach an equilibrium between phosphorylated and un-phosphorylated states of the protein substrate, and the fluorescence characteristics will reflect this equilibrium level. In one aspect, this method can be used to compare mutant cells to identify which ones possess greater or lesser ratio of kinase to phosphatase activity. Such cells can be sorted by a fluorescent cell sorter based on fluorescence.

A contemplated variation of the above assay is to use the controlling nucleotide sequences to produce a sudden increase in the expression of either the fluorescent protein substrate or the enzyme being assayed, e.g., by inducing expression of the construct. A fluorescent property is monitored at one or more time intervals after the onset of increased expression. A high amount of the property associated with phosphorylated state reflects a large amount or high efficiency of the kinase. This kinetic determination has the advantage of minimizing any dependency of the assay on the rates of degradation or loss of the fluorescent protein moieties.

In another embodiment, the vector may be incorporated into an entire organism by standard transgenic or gene replacement techniques. An expression vector capable of expressing the enzyme optionally may be incorporated into the entire organism by standard transgenic or gene replacement techniques. Then, a sample from the organism containing the protein substrate is tested. For example, cell or tissue homogenates, individual cells, or samples of body fluids, such as blood, can be tested.

II. SCREENING ASSAYS

The enzymatic assays of the invention can be used in drug screening assays to determine whether a compound alters the activity of a protein kinase. In one embodiment, the assay is performed on a sample *in vitro* containing the enzyme. A sample containing a known amount of enzyme activity is mixed with a substrate of the invention and with a test compound. The amount of the enzyme activity in the sample is then determined as above, e.g., by measuring the amount of a fluorescent property at a first and second time after contact between the sample, the protein substrate, a phosphate substrate, and the compound. Then the amount of activity per mole of enzyme in the presence of the test compound is compared with the activity per mole of enzyme in the absence of the test compound. A difference indicates that the test compound alters the activity of the enzyme.

In another embodiment, the ability of a compound to alter kinase activity *in vivo* is determined. In an *in vivo* assay, cells transfected with a expression vector encoding a substrate of the invention are exposed to different amounts of the test compound, and the effect on fluorescence in each cell can be determined. Typically, the difference is calibrated against standard measurements to yield an absolute amount of kinase activity. A test compound that inhibits or blocks the activity or expression of the

kinase can be detected by a relative increase in the property associated with the un-phosphorylated state. The cell can also be transfected with an expression vector to co-express the kinase or an upstream signaling component such as a receptor, and fluorescent substrate. This method is useful for detecting signaling to a protein kinase of interest from an upstream component of the signaling pathway. If a signal from an upstream molecule, e.g., a receptor, is inhibited by a drug activity, then the kinase activity will not be altered from basal. This provides a method for screening for compounds which affect cellular events (including receptor-ligand binding, protein-protein interactions or kinase activation) which signal to the target kinase.

This invention also provides kits containing the fluorescent protein substrate and a phosphate substrate for the protein kinase. In one embodiment, the kit has a container holding the fluorescent protein substrate and another container holding the phosphate substrate. Protein kinases of known activity could be included for use as positive controls and standards.

III. FLUORESCENT PROTEIN SUBSTRATES FOR PROTEIN KINASES

As used herein, the term "fluorescent property" refers to the molar extinction coefficient at an appropriate excitation wavelength, the fluorescence quantum efficiency, the shape of the excitation spectrum or emission spectrum, the excitation wavelength maximum and emission wavelength maximum, the ratio of excitation amplitudes at two different wavelengths, the ratio of emission amplitudes at two different wavelengths, the excited state lifetime, or the fluorescence anisotropy. A measurable difference in any one of these properties between the phosphorylated and un-phosphorylated states suffices for the utility of the fluorescent protein substrates of the invention in assays for kinase activity. A measurable difference can be determined by determining the amount of any quantitative fluorescent property, e.g., the amount of fluorescence at a particular wavelength, or the integral of fluorescence over the emission spectrum. Optimally, the protein substrates are selected to have fluorescent properties that are easily distinguishable in the un-phosphorylated and phosphorylated states. Determining ratios of excitation amplitude or emission amplitude at two different wavelengths ("excitation amplitude ratioing" and "emission amplitude ratioing", respectively) are particularly advantageous because the ratioing process provides an internal reference and cancels out variations in the absolute brightness of the excitation

source, the sensitivity of the detector, and light scattering or quenching by the sample. Furthermore, if phosphorylation of the protein substrate changes its ratio of excitation or emission amplitudes at two different wavelengths, then such ratios measure the extent of phosphorylation independent of the absolute quantity of the protein substrate. Some of the fluorescent protein substrates described herein do exhibit a phosphorylation-induced change in the ratio of excitation amplitudes at two different wavelengths. Even if a fluorescent protein substrate does not exhibit a phosphorylation-induced change in excitation or emission amplitudes at two wavelengths, cells can be provided that co-express another fluorescent protein that is not sensitive to phosphorylation and whose excitation or emission spectrum is peaked at wavelengths distinct from those of the phosphorylation substrate. Provided that the expression of the two proteins are both controlled by the same nucleotide control sequences, their expression levels should be closely linked. Therefore ratioing the excitation or emission amplitude of the phosphorylation substrate at its preferred wavelength to the corresponding excitation or emission amplitude of the phosphorylation-insensitive reference protein at its separate preferred wavelength is an alternative method for canceling out variations in the absolute quantity of cells or overall level of protein expression.

A. Fluorescent Proteins

As used herein, the term "fluorescent protein" refers to any protein capable of fluorescence when excited with appropriate electromagnetic radiation. This includes fluorescent proteins whose amino acid sequences are either naturally occurring or engineered (i.e., analogs). Many cnidarians use green fluorescent proteins ("GFPs") as energy-transfer acceptors in bioluminescence. A "green fluorescent protein," as used herein, is a protein that fluoresces green light. Similarly, "blue fluorescent proteins" fluoresce blue light and "red fluorescent proteins" fluoresce red light. GFPs have been isolated from the Pacific Northwest jellyfish, *Aequorea victoria*, the sea pansy, *Renilla reniformis*, and *Phialidium gregarium*. W.W. Ward et al., *Photochem. Photobiol.*, 35:803-808 (1982); L.D. Levine et al., *Comp. Biochem. Physiol.*, 72B:77-85 (1982).

A variety of *Aequorea*-related fluorescent proteins having useful excitation and emission spectra have been engineered by modifying the amino acid sequence of a naturally occurring GFP from *Aequorea victoria*. (D.C. Prasher et al., *Gene*, 111:229-233 (1992); R. Heim et al., *Proc. Natl. Acad. Sci., USA*, 91:12501-04 (1994); U.S.

patent application 08/337,915, filed November 10, 1994; International application PCT/US95/14692, filed 11/10/95.)

As used herein, a fluorescent protein is an "*Aequorea*-related fluorescent protein" if any contiguous sequence of 150 amino acids of the fluorescent protein has at least 85% sequence identity with an amino acid sequence, either contiguous or non-contiguous, from the 238 amino-acid wild-type *Aequorea* green fluorescent protein of Fig. 3 (SEQ ID NO:2). More preferably, a fluorescent protein is an *Aequorea*-related fluorescent protein if any contiguous sequence of 200 amino acids of the fluorescent protein has at least 95% sequence identity with an amino acid sequence, either contiguous or non-contiguous, from the wild type *Aequorea* green fluorescent protein of Fig. 3 (SEQ ID NO:2). Similarly, the fluorescent protein may be related to *Renilla* or *Phialidium* wild-type fluorescent proteins using the same standards.

Optimal alignment of sequences for aligning a comparison window may be conducted by the local homology algorithm of Smith and Waterman (1981) *Adv. Appl. Math.*, 2:482, by the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.*, 48:443, by the search for similarity method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci., U.S.A.*, 85:2444, by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Dr., Madison, WI), or by inspection. The best alignment (i.e., resulting in the highest percentage of homology over the comparison window, i.e., 150 or 200 amino acids) generated by the various methods is selected.

The percentage of sequence identity is calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical amino acid occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison (i.e., the window size), and multiplying the result by 100 to yield the percentage of sequence identity.

Aequorea-related fluorescent proteins include, for example and without limitation, wild-type (native) *Aequorea victoria* GFP (D.C. Prasher et al., "Primary structure of the *Aequorea victoria* green fluorescent protein," *Gene*, (1992) 111:229-33), whose nucleotide sequence (SEQ ID NO:1) and deduced amino acid sequence (SEQ ID NO:2) are presented in Fig. 3, allelic variants of this sequence, e.g., Q80R, which has

the glutamine residue at position 80 substituted with arginine (M. Chalfie et al., *Science*, (1994) 263:802-805), those *Aequorea*-related engineered versions described in Table I, variants that include one or more folding mutations and fragments of these proteins that are fluorescent, such as *Aequorea* green fluorescent protein from which the two amino-terminal amino acids have been removed. Several of these contain different aromatic amino acids within the central chromophore and fluoresce at a distinctly shorter wavelength than wild type species. For example, mutants P4 and P4-3 contain (in addition to other mutations) the substitution Y66H, whereas W2 and W7 contain (in addition to other mutations) Y66W. Other mutations both close to the chromophore region of the protein and remote from it in primary sequence may affect the spectral properties of GFP and are listed in the first part of the table below.

TABLE I

	<u>Clone</u>	<u>Mutation(s)</u>	<u>Excitation max (nm)</u>	<u>Emission max (nm)</u>	<u>Extinct. Coeff. (M⁻¹cm⁻¹)</u>	<u>Quantum yield</u>
5	Wild type	none	395 (475)	508	21,000 (7,150)	0.77
	P4	Y66H	383	447	13,500	0.21
	P4-3	Y66H Y145F	381	445	14,000	0.38
	W7	Y66W N146I M153T V163A N212K	433 (453)	475 (501)	18,000 (17,100)	0.67
	W2	Y66W I123V Y145H H148R M153T V163A N212K	432 (453)	480	10,000 (9,600)	0.72
10	S65T	S65T	489	511	39,200	0.68
	P4-1	S65T M153A K238E	504 (396)	514	14,500 (8,600)	0.53
	S65A	S65A	471	504		
	S65C	S65C	479	507		
	S65L	S65L	484	510		
15	Y66F	Y66F	360	442		
	Y66W	Y66W	458	480		

20

Additional mutations in *Aequorea*-related fluorescent proteins, referred to as "folding mutations," improve the ability of GFP to fold at higher temperatures, and to be more fluorescent when expressed in mammalian cells, but have little or no effect on the peak wavelengths of excitation and emission. It should be noted that these may be

combined with mutations that influence the spectral properties of GFP to produce proteins with altered spectral and folding properties. Folding mutations include: T44A, F64L, V68L, S72A, F99S, Y145F, N146I, M153T or A, V163A, I167T, S175G, S205T and N212K.

5 This invention contemplates the use of other fluorescent proteins in fluorescent protein substrates for protein kinases. The cloning and expression of yellow fluorescent protein from *Vibrio fischeri* strain Y-1 has been described by T.O. Baldwin et al., *Biochemistry* (1990) 29:5509-15. This protein requires flavins as fluorescent co-factors. The cloning of Peridinin-chlorophyll *a* binding protein from the dinoflagellate
10 *Symbiodinium* sp. was described by B.J. Morris et al., *Plant Molecular Biology*, (1994) 24:673:77. One useful aspect of this protein is that it fluoresces red. The cloning of phycobiliproteins from marine cyanobacteria such as *Synechococcus*, e.g., phycoerythrin and phycocyanin, is described in S.M. Wilbanks et al., *J. Biol. Chem.* (1993) 268:1226-35. These proteins require phycobilins as fluorescent co-factors, whose insertion into the
15 proteins involves auxiliary enzymes. The proteins fluoresce at yellow to red wavelengths.

 As used herein, the "fluorescent protein moiety" of a fluorescent protein substrate is that portion of the amino acid sequence of a fluorescent protein substrate which, when the amino acid sequence of the fluorescent protein substrate is optimally
20 aligned with the amino acid sequence of a naturally occurring fluorescent protein, lies between the amino terminal and carboxy terminal amino acids, inclusive, of the amino acid sequence of the naturally occurring fluorescent protein.

 It has been found that fluorescent proteins can be genetically fused to other target proteins and used as markers to identify the location and amount of the target
25 protein produced. Accordingly, this invention provides fusion proteins comprising a fluorescent protein moiety and additional amino acid sequences. Such sequences can be, for example, up to about 15, up to about 50, up to about 150 or up to about 1000 amino acids long. The fusion proteins possess the ability to fluoresce when excited by electromagnetic radiation. In one embodiment, the fusion protein comprises a
30 polyhistidine tag to aid in purification of the protein.

B. Phosphorylation Sites For Protein Kinases

Fluorescent protein substrates for a protein kinase are the subset of fluorescent proteins as defined above whose amino acid sequence includes a phosphorylation site. Fluorescent protein substrates can be made by modifying the amino acid sequence of an existing fluorescent protein to include a phosphorylation site for a protein kinase. Fluorescent protein substrates for protein kinases are not meant to include isolated fluorescent proteins that have a naturally occurring phosphorylation site, naturally occurring fluorescent proteins or currently known mutant fluorescent proteins. Such previously known fluorescent proteins or mutants may be substrates for protein kinases, but do not exhibit any detectable change in fluorescent properties upon phosphorylation.

As used herein, the term "phosphorylation site for a protein kinase" refers to an amino acid sequence which, as part of a polypeptide, is recognized by a protein kinase for the attachment of a phosphate moiety. The phosphorylation site can be a site recognized by, for example, protein kinase A, a cGMP-dependent protein kinase, protein kinase C, Ca^{2+} /calmodulin-dependent protein kinase I, Ca^{2+} /calmodulin-dependent protein kinase II or MAP kinase activated protein kinase type 1.

The preferred consensus sequence for protein kinase A is RRXSZ (SEQ ID NO:3) or RRXTZ (SEQ ID NO:4), wherein X is any amino acid and Z is a hydrophobic amino acid, preferably valine, leucine or isoleucine. Many variations in the above sequence are allowed, but generally exhibit poorer kinetics. For example, lysine (K) can be substituted for the second arginine. Many consensus sequences for other protein kinases have been tabulated, e.g. by Kemp, B.E. and Pearson, R.B. (1990) *Trends Biochem. Sci.* 15: 342-346; Songyang, Z. et al. (1994) *Current Biology* 4: 973-982.

For example, a fluorescent protein substrate selective for phosphorylation by cGMP-dependent protein kinase can include the following consensus sequence: BKISASEFDR PLR (SEQ ID NO:5), where B represents either lysine (K) or arginine (R), and the first S is the site of phosphorylation (Colbran et al. (1992) *J. Biol. Chem.* 267: 9589-9594). The residues DRPLR (SEQ ID NO:6) are less critical than the phenylalanine (F) just preceding them for specific recognition by cGMP-dependent protein kinase in preference to cAMP-dependent protein kinase.

Either synthetic or naturally occurring motifs can be used to create a protein kinase phosphorylation site. For example, peptides including the motif XRXSXR (SEQ ID NO:7), wherein X is any amino acid, are among the best synthetic substrates (Kemp and Pearson, *supra*) for protein kinase C. Alternatively, the Myristoylated Alanine-Rich Kinase C substrate ("MARCKS") is one of the best substrates for PKC and is a real target for the kinase *in vivo*. The sequence around the phosphorylation site of MARCKS is KKKRFSFK (SEQ ID NO:8) (Graff et al. (1991) *J. Biol. Chem.* 266:14390-14398). Either of these two sequences can be incorporated into a fluorescent protein to make it a substrate for protein kinase C.

A protein substrate for Ca^{2+} /calmodulin-dependent protein kinase I is derived from the sequence of synapsin I, a known optimal substrate for this kinase. The recognition sequence around the phosphorylation site is LRRLSDSNF (SEQ ID NO:9) (Lee et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:6413-6417).

A protein substrate selective for Ca^{2+} /calmodulin-dependent protein kinase II is derived from the sequence of glycogen synthase, a known optimal substrate for this kinase. The recognition sequence around the phosphorylation site is KKLNRSLTVA (SEQ ID NO:10) (Stokoe et al. (1993) *Biochem. J.* 296:843-849). A small change in this sequence to KKANRTLSVA (SEQ ID NO:11) makes the latter specific for MAP kinase activated protein kinase type 1.

In one embodiment, the fluorescent protein substrate contains a phosphorylation site around one of the termini, in particular, the amino-terminus, of the fluorescent protein moiety. The site preferably is located in a position within five, ten, fifteen, or twenty amino acids of a position corresponding to the wild type amino-terminal amino acid of the fluorescent protein moiety ("within twenty amino acids of the amino-terminus"). This includes sites engineered into the existing amino acid sequence of the fluorescent protein moiety and sites produced by extending the amino terminus of the fluorescent protein moiety.

One may, for example, modify the existing sequence of wild type *Aequorea* GFP or a variant or it as listed above to include a phosphorylation site within the first ten or twenty amino acids. In one embodiment, the naturally occurring sequence is modified as follows:

wild type: MSKGEELFTG (1-10 of SEQ ID NO:2)

substrate: MRRRRSIITG (SEQ ID NO:12).

One may include modifying the naturally occurring sequence of *Aequorea* GFP by introducing a phosphorylation site into an extended amino acid sequence of such a protein created by adding flanking sequences to the amino terminus, for example:

wild type: MSKGEELFTG (1-10 of SEQ ID NO:2)

substrate: MRRRRSIIIFTG (SEQ ID NO:13).

Fluorescent protein substrates having a phosphorylation site around a terminus of the fluorescent protein moiety offer the following advantages. First, it is often desirable to append additional amino acid residues onto the fluorescent protein moiety in order to create a specific phosphorylation consensus sequence. Such a sequence is much less likely to disrupt the folding pattern of the fluorescent protein when appended onto the terminus than when inserted into the interior of the protein sequence. Second, different phosphorylation motifs can be interchanged without significant disruption of GFP therefore providing a general method of measuring different kinases. Third, the phosphorylation site is exposed to the surface of the protein and, therefore, more accessible to protein kinases. Fourth, we have discovered that phosphorylation at sites close to the N-terminus of GFP can provide large changes in fluorescent properties if the site of phosphorylation is chosen such that the Ser or Thr residue which is phosphorylated occupies a position which in the wild-type protein was originally negatively or positively charged. Specifically, replacement of Glu 5 or Glu 6 by a non-charged Ser or Thr residue can significantly disrupt fluorescence of GFP when made within the right context of surrounding amino acids. Phosphorylation of the serine or threonine will restore negative charge to this position and thereby increases fluorescence.

In another embodiment, the fluorescent protein substrate includes a phosphorylation site remote from the terminus, e.g., that is separated by more than about twenty amino acids from the terminus of the fluorescent protein moiety and within the fluorescent protein moiety. One embodiment of this form includes the *Aequorea*-related fluorescent protein substrate comprising the substitution H217S, creating a consensus protein kinase A phosphorylation site. Additionally, phosphorylation sites comprising the following alterations based on the sequence of wild type *Aequorea* GFP exhibit

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fluorescent changes upon phosphorylation: 69RRFSA (SEQ ID NO:14) and 214KRDSM (SEQ ID NO:15).

The practitioner should consider the following in selecting amino acids for substitution within the fluorescent protein moiety remote in primary amino acid sequence from the terminus. First, it is preferable to select amino acid sequences within the fluorescent protein moiety that resemble the sequence of the phosphorylation site. In this way, fewer amino acid substitutions in the native protein are needed to introduce the phosphorylation site into the fluorescent protein. For example, protein kinase A recognizes the sequence RRXSZ (SEQ ID NO:3) or RRXTZ (SEQ ID NO:4), wherein X is any amino acid and Z is a hydrophobic amino acid. Serine or threonine is the site of phosphorylation. It is preferable to introduce this sequence into the fluorescent protein moiety at sequences already containing Ser or Thr, so that Ser or Thr are not substituted in the protein. More preferably the phosphorylation site is created at locations having some existing homology to the sequence recognized by protein kinase A, e.g., having a proximal Arg or hydrophobic residues with the same spatial relationship as in the phosphorylation site.

Second, locations on the surface of the fluorescent protein are preferred for phosphorylation sites. This is because surface locations are more likely to be accessible to protein kinases than interior locations. Surface locations can be identified by computer modeling of the fluorescent protein structure or by reference to the crystal structure of *Aequorea* GFP. Also, charged amino acids in the fluorescent protein are more likely to lie on the surface than inside the fluorescent protein, because such amino acids are more likely to be exposed to water in the environment.

In cases where the phosphorylation site is either at the N-terminus or remote from it, the amino acid context around the phosphorylation site needs to be optimized in order to maximize the change in fluorescence. Amino acid substitutions that change large bulky and or hydrophobic amino acids to smaller and less hydrophobic replacements are generally helpful. Similarly large charged amino acids can be replaced by smaller, less charged amino acids. For example:

a/Hydrophobic to less hydrophobic

Phe to Leu

Leu to Ala

b/Charged to charged but smaller

Glu to Asp

Arg to Lys

c/Charged to less charged

Glu to Gln

Asp to Asn

d/Charged to polar

Glu to Thr

Asp to Ser

e/Charged to non-polar

Glu to Leu

Asp to Ala

These changes can be accomplished by directed means or using random iterative approaches where changes are made randomly and the best ones selected based upon their change in fluorescent properties after phosphorylation by an appropriate kinase.

Third, amino acids at distant locations from the actual site of phosphorylation can be varied to enhance fluorescence changes upon phosphorylation. These mutations can be created through site directed mutagenesis, or through random mutagenesis, for example by error-prone PCR, to identify mutations that enhance either absolute fluorescence or the change in fluorescence upon phosphorylation. The identification of mutants remote in primary sequence from the N-terminus identifies potentially interacting sequences which may provide additional areas in which further mutagenesis could be used to refine the change in fluorescence upon phosphorylation. For example, it has been determined that mutations around the amino terminus phosphorylation site interact (either transiently during folding, or in a stable fashion) with amino acids at positions 171 and 172, and that point mutations that significantly disrupt

fluorescence of GFP by changing negative to positive charges near the amino terminus can be rescued by changing a positive to a negative charge at position 171.

In the phosphorylation mutant 50 the sequence is a/ and for reference the wild type sequence b/ is listed below.

5

a/ MSKRRDSLT (SEQ ID NO:16)

b/ MSKGEELFT (1-9 of SEQ ID NO:2)

10 The phosphorylation mutant has only 7% of the fluorescence of wild type protein. However, its fluorescence can be restored to 80% of wild type by 2 amino acid changes, E171K and I172V, positions which are quite remote in linear sequence from the amino terminus.

15 Thus, changes in charge at E171K (negative to positive) can almost completely restore the fluorescence of the phosphorylation mutant, strongly suggesting that the original loss of fluorescence arose primarily through changes in charge caused by the point mutations. It is clear that the addition and loss of charge at positions around, and at the phosphorylation site, have a significant impact on fluorescence formation. The fact that charge alone can significantly affect the fluorescence properties of GFP is highly significant within the scope of the present application since phosphorylation involves the
20 addition of 2 negative charges associated with the phosphate group (OPO_3^{3-}) on the serine residue.

In the above case the mutations restore fluorescence of the phosphorylation mutant, without significantly increasing the magnitude of the change in fluorescence upon phosphorylation. Nevertheless the identification of these positions in GFP provides a
25 valuable tool to further optimize changes in fluorescence upon phosphorylation by creating random mutations at codons around positions 171, 172 and 173 to identify mutations that enhance changes in fluorescence upon phosphorylation.

This can be achieved by co-expressing the kinase of interest with the fluorescent substrate of the invention containing random mutations which may enhance
30 the fluorescence changes upon phosphorylation in bacteria (in the example above these would be NNK mutations at codons 171, 172 and 173, where N represents a random choice of any of the four bases and K represents a random choice of guanine or

thymine). The expression vector containing the mutated fluorescent substrates and the kinase are transformed into host bacteria and the individual bacterial colonies grown up. Each colony is derived from a single cell, and hence contains a single unique mutant fluorescent substrate grown up.

5 The individual colonies may then be grown up and screened for fluorescence either by fluorescence activated cell sorting (FACS), or by observation under a microscope. Those that exhibit the greatest fluorescence can then be rescreened under conditions in which the kinase gene is inactivated. This can be achieved by appropriate digests of the kinase gene by restriction enzymes that specifically cut within
10 the kinase but not GFP. Comparison of the brightness of the mutant first in the presence of kinase then in its absence indicates the relative effect of phosphorylation on the mutant GFP.

C. Production Of Fluorescent Protein Substrates For Protein Kinases

15 While certain fluorescent protein substrates for protein kinases can be prepared chemically, for example, by coupling a peptide moiety to the amino terminus of a fluorescent protein, it is preferable produce fluorescent protein substrates recombinantly.

 Recombinant production of a fluorescent protein substrate involves
20 expressing a nucleic acid molecule having sequences that encode the protein. As used herein, the term "nucleic acid molecule" includes both DNA and RNA molecules. It will be understood that when a nucleic acid molecule is said to have a DNA sequence, this also includes RNA molecules having the corresponding RNA sequence in which "U" replaces "T." The term "recombinant nucleic acid molecule" refers to a nucleic acid
25 molecule which is not naturally occurring, and which comprises two nucleotide sequences which are not naturally joined together. Recombinant nucleic acid molecules are produced by artificial combination, e.g., genetic engineering techniques or chemical synthesis.

 In one embodiment, the nucleic acid encodes a fusion protein in which a
30 single polypeptide includes the fluorescent protein moiety within a longer polypeptide. In another embodiment the nucleic acid encodes the amino acid sequence of consisting

essentially of a fluorescent protein modified to include a phosphorylation site. In either case, nucleic acids that encode fluorescent proteins are useful as starting materials.

Nucleic acids encoding fluorescent proteins can be obtained by methods known in the art. For example, a nucleic acid encoding a green fluorescent protein can be isolated by polymerase chain reaction of cDNA from *A. victoria* using primers based on the DNA sequence of *A. victoria* green fluorescent protein, as presented in Fig. 3. PCR methods are described in, for example, U.S. Pat. No. 4,683,195; Mullis et al. (1987) *Cold Spring Harbor Symp. Quant. Biol.* 51:263; and Erlich, ed., *PCR Technology*, (Stockton Press, NY, 1989).

Mutant versions of fluorescent proteins can be made by site-specific mutagenesis of other nucleic acids encoding fluorescent proteins, or by random mutagenesis caused by increasing the error rate of PCR of the original polynucleotide with 0.1 mM MnCl₂ and unbalanced nucleotide concentrations. See, e.g., U.S. patent application 08/337,915, filed November 10, 1994 or International application PCT/US95/14692, filed 11/10/95.

Nucleic acids encoding fluorescent protein substrates which are fusions between a polypeptide including a phosphorylation site and a fluorescent protein and can be made by ligating nucleic acids that encode each of these. Nucleic acids encoding fluorescent protein substrates which include the amino acid sequence of a fluorescent protein in which one or more amino acids in the amino acid sequence of a fluorescent protein are substituted to create a phosphorylation site can be created by, for example, site specific mutagenesis of a nucleic acid encoding a fluorescent protein.

The construction of expression vectors and the expression of genes in transfected cells involves the use of molecular cloning techniques also well known in the art. Sambrook et al., *Molecular Cloning -- A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, (1989) and *Current Protocols in Molecular Biology*, F.M. Ausubel et al., eds., (Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc.

Nucleic acids used to transfect cells with sequences coding for expression of the polypeptide of interest generally will be in the form of an expression vector including expression control sequences operatively linked to a nucleotide sequence coding for expression of the polypeptide. As used, the term nucleotide sequence "coding for

expression of" a polypeptide refers to a sequence that, upon transcription and translation of mRNA, produces the polypeptide. As any person skilled in the art recognizes, this includes all degenerate nucleic acid sequences encoding the same amino acid sequence.

This can include sequences containing, e.g., introns. As used herein, the term

5 "expression control sequences" refers to nucleic acid sequences that regulate the expression of a nucleic acid sequence to which it is operatively linked. Expression control sequences are "operatively linked" to a nucleic acid sequence when the expression control sequences control and regulate the transcription and, as appropriate, translation of the nucleic acid sequence. Thus, expression control sequences can include
10 appropriate promoters, enhancers, transcription terminators, a start codon (i.e., ATG) in front of a protein-encoding gene, splicing signals for introns, maintenance of the correct reading frame of that gene to permit proper translation of the mRNA, and stop codons.

The recombinant nucleic acid can be incorporated into an expression vector comprising expression control sequences operatively linked to the recombinant
15 nucleic acid. The expression vector can be adapted for function in prokaryotes or eukaryotes by inclusion of appropriate promoters, replication sequences, markers, etc.

The expression vector can be transfected into a host cell for expression of the recombinant nucleic acid. Host cells can be selected for high levels of expression in order to purify the protein. *E. coli* is useful for this purpose. Alternatively, the host
20 cell can be a prokaryotic or eukaryotic cell selected to study the activity of an enzyme produced by the cell. The cell can be, e.g., a cultured cell or a cell *in vivo*.

Recombinant fluorescent protein substrates can be produced by expression of nucleic acid encoding for the protein in *E. coli*. *Aequorea*-related fluorescent proteins are best expressed by cells cultured between about 15° C and 30° C but higher
25 temperatures (e.g. 37° C) are possible. After synthesis, these enzymes are stable at higher temperatures (e.g., 37° C) and can be used in assays at those temperatures.

The construct can also contain a tag to simplify isolation of the substrate. For example, a polyhistidine tag of, e.g., six histidine residues, can be incorporated at the amino or carboxyl terminal of the fluorescent protein substrate. The polyhistidine tag
30 allows convenient isolation of the protein in a single step by nickel-chelate chromatography.

Alternatively, the substrates need not be isolated from the host cells. This method is particularly advantageous for the assaying for the presence of protein kinase activity *in situ*.

5 IV. LIBRARIES OF CANDIDATE SUBSTRATES

 The inclusion of a phosphorylation site around the amino terminus of a fluorescent protein moiety can provide a fluorescent protein that, when phosphorylated, can alter a fluorescent property of the protein. Accordingly, this invention provides libraries of fluorescent protein candidate substrates useful for screening in the
10 identification and characterization of sequences that can be recognized and efficiently phosphorylated by a kinase. Libraries of these proteins can be screened to identify sequences that can be phosphorylated by kinases of unknown substrate specificity, or to characterize differences in kinase activity in, or from, diseased and normal cells or tissues.

15 As used herein, a "library" refers to a collection containing at least 10 different members. Each member of a fluorescent protein candidate substrate library comprises a fluorescent protein moiety and a variable peptide moiety, which is preferably located near the amino-terminus of the fluorescent protein moiety and preferably has fewer than about 15 amino acids. The variety of amino acid sequences for the peptide
20 moiety is at the discretion of the practitioner. For example, the library can contain a quite diverse collection of variable peptide moieties in which most or all of the amino acid positions are subjected to a non-zero but low probability of substitution. Also, the library can contain variable peptide moieties having an amino acid sequence in which only a few, e.g., one to ten, amino acid positions are varied, but the probability of
25 substitution at each position is relatively high.

 Preferably, libraries of fluorescent protein candidate substrates are created by expressing protein from libraries of recombinant nucleic acid molecules having expression control sequences operatively linked to nucleic acid sequences that code for the expression of different fluorescent protein candidate substrates. Methods of making
30 nucleic acid molecules encoding a diverse collection of peptides are described in, for example, U.S. patent 5,432,018 (Dower et al.), U.S. patent 5,223,409 (Ladner et al.), U.S. Patent 5,264,563 (Huse), and International patent publication WO 92/06176 (Huse

et al.). For expression of fluorescent protein candidate substrates, recombinant nucleic acid molecules are used to transfect cells, such that each cell contains a member of the library. This produces, in turn, a library of host cells capable of expressing the library of different fluorescent protein candidate substrates. The library of host cells is useful in the screening methods of this invention.

In one method of creating such a library, a diverse collection of oligonucleotides having preferably random codon sequences are combined to create polynucleotides encoding peptides having a desired number of amino acids. The oligonucleotides preferably are prepared by chemical synthesis. The polynucleotides encoding variable peptide moiety can then be coupled to the 5' end of a nucleic acid coding for the expression of a fluorescent protein moiety or a carboxy-terminal portion of it. That is, the fluorescent protein moiety can be cut back to eliminate up to 20 amino acids of the reference fluorescent protein. This creates a recombinant nucleic acid molecule coding for the expression of a fluorescent protein candidate substrate having a peptide moiety fused to the amino terminus of the fluorescent protein. This recombinant nucleic acid molecule is then inserted into an expression vector to create a recombinant nucleic acid molecule comprising expression control sequences operatively linked to the sequences encoding the candidate substrate.

To generate the collection of oligonucleotides which forms a series of codons encoding a random collection of amino acids and which is ultimately cloned into the vector, a codon motif is used, such as $(NNK)_x$, where N may be A, C, G, or T (nominally equimolar), K is G or T (nominally equimolar), and x is the desired number of amino acids in the peptide moiety, e.g., 15 to produce a library of 15-mer peptides. The third position may also be G or C, designated "S". Thus, NNK or NNS (i) code for all the amino acids, (ii) code for only one stop codon, and (iii) reduce the range of codon bias from 6:1 to 3:1. The expression of peptides from randomly generated mixtures of oligonucleotides in appropriate recombinant vectors is discussed in Oliphant et al., *Gene* 44:177-183 (1986).

An exemplified codon motif $(NNK)_6$ (SEQ ID NO:17) produces 32 codons, one for each of 12 amino acids, two for each of five amino acids, three for each of three amino acids and one (amber) stop codon. Although this motif produces a codon

distribution as equitable as available with standard methods of oligonucleotide synthesis, it results in a bias against peptides containing one-codon residues.

An alternative approach to minimize the bias against one-codon residues involves the synthesis of 20 activated tri-nucleotides, each representing the codon for one of the 20 genetically encoded amino acids. These are synthesized by conventional means, removed from the support but maintaining the base and 5-HO-protecting groups, and activating by the addition of 3'O-phosphoramidite (and phosphate protection with beta-cyanoethyl groups) by the method used for the activation of mononucleosides, as generally described in McBride and Caruthers, *Tetrahedron Letters* 22:245 (1983).

Degenerate "oligocodons" are prepared using these trimers as building blocks. The trimers are mixed at the desired molar ratios and installed in the synthesizer. The ratios will usually be approximately equimolar, but may be a controlled unequal ratio to obtain the over- to under-representation of certain amino acids coded for by the degenerate oligonucleotide collection. The condensation of the trimers to form the oligocodons is done essentially as described for conventional synthesis employing activated mononucleosides as building blocks. See generally, Atkinson and Smith, *Oligonucleotide Synthesis*, M.J. Gait, ed. p35-82 (1984). Thus, this procedure generates a population of oligonucleotides for cloning that is capable of encoding an equal distribution (or a controlled unequal distribution) of the possible peptide sequences.

Libraries of amino terminal phosphorylation sites may also be annealed to libraries of randomly mutated GFP sequences to enable the selection of optimally responding substrates.

V. METHODS FOR SCREENING LIBRARIES OF CANDIDATE SUBSTRATES

Libraries of host cells expressing fluorescent protein candidate substrates are useful in identifying fluorescent proteins having peptide moieties that alter a fluorescent property of the fluorescent protein. Several methods of using the libraries are envisioned. In general, one begins with a library of recombinant host cells, each of which expresses a different fluorescent protein candidate substrate. Each cell is expanded into a clonal population that is genetically homogeneous.

In a first method, the desired fluorescent property is measured from each clonal population before and at least one specified time after a known change in

intracellular protein kinase activity. This change in kinase activity could be produced by transfection with a gene encoding the kinase, by induction of kinase gene expression using expression control elements, or by any condition that post-translationally modulates activity of a kinase that has already been expressed. Examples of the latter include cell surface receptor mediated elevation of intracellular cAMP to activate cAMP-dependent surface receptor mediated increases of intracellular cGMP to activate cGMP-dependent protein kinase, cytosolic free calcium to activate Ca^{2+} /calmodulin-dependent protein kinase types I, II, or IV, or the production of diacylglycerol to activate protein kinase C, etc. One then selects for the clone(s) that show the biggest or fastest change in the desired fluorescence property. This method detects fluorescent protein mutants whose folding and maturation was influenced by phosphorylation as well as those affected by phosphorylation after maturation.

One embodiment of this method exploits the fact that the catalytic subunit of cAMP-dependent protein kinase is constitutively active in the absence of the regulatory subunit and is growth-inhibitory in *E. coli* and most mammalian cells. Therefore, the cells tend to shed the kinase gene by recombination. The change in kinase activity is obtained by culturing the cells for a time sufficient to lose the kinase gene.

In a second method the host cells do not express the protein kinase of interest. Each clonal population is separately lysed. ATP is then added to the lysate. After an incubation period to allow phosphorylation by background kinases, the fluorescence property is measured. Then exogenous protein kinase is added to the lysate and the fluorescent property is re-measured at one or more specified time points. Again one selects for the clone(s) that show the biggest or fastest change(s) in the desired fluorescence property. Because little or no fresh protein synthesis is likely to occur in the lysate, this method would discriminate against mutants which are sensitive to phosphorylation only during their folding and maturation.

In one embodiment of this method, the lysate is split into two aliquots, one of which is mixed with kinase and ATP, the other of which receives only ATP. One selects for the clone(s) that show the biggest difference in fluorescence property between the two aliquots.

The nucleic acids from cells exhibiting the different properties can be isolated from the cells. Candidate substrates having different fluorescent properties can be tested further to identify the source of the difference.

5 The host cell also can be transfected with an expression vector capable of expressing an enzyme, such as a protein kinase, whose effect on the fluorescent property is to be tested.

VI. EXAMPLES

10 A. Phosphorylation sites located in the amino acid sequence of *Aequorea* GFP remote in the primary amino acid sequence from the N-terminus

Potential sites for phosphorylation were chosen at or close to positions in GFP which had previously been identified to exert significant effects on fluorescence, or which had a higher probability of surface exposure based on computer algorithms (Fig. 4). For example, in a mutant called H9, Ser202 and Thr203 are mutated to F and I
15 respectively, creating a large change in spectral properties (see also Ehrig *et al*, 1995). Therefore in one mutant, 199RRLSI (SEQ ID NO:18), a potential site of phosphorylation was created around Ser202, whose phosphorylation should significantly affect the fluorescent properties. Similarly the amino acids located at positions 72 and 175 have been implicated in increased folding efficiency of GFP at higher temperatures and were
20 made into potential sites of phosphorylation in separate mutants.

A complete list of the positions and amino acid changes made for each phosphorylation mutant in this series is outlined in Fig. 4. GFP was expressed in *E. coli* using the expression plasmid pRSET (Invitrogen), in which the region encoding GFP was fused in frame with nucleotides encoding an N-terminal polyhistidine tag (Fig. 5). The
25 sequence changes were introduced by site-directed mutagenesis using the Bio-Rad mutagenesis kit (Kunkel, T.A. (1985) *Proc. Natl. Acad. Sci.* 82:488-492, Kunkel, T.A., Roberts, J.D., and Zakour, R.A. (1987) *Meth Enzymol* 154:367-382) and confirmed by sequencing. The recombinant proteins were induced with IPTG and expressed in bacteria and purified by nickel affinity chromatography. The sequence changes, relative
30 fluorescence, relative rate of phosphorylation and % change in fluorescence upon phosphorylation are listed in Table II. In those cases where the protein exhibited no

fluorescence after insertion of the phosphorylation site no determinations were made on the effect of phosphorylation on fluorescence.

Table II: Relative fluorescence, rate of phosphorylation and change in fluorescence upon phosphorylation for mutants incorporating phosphorylation sites remote from the N-terminus

SEQ ID NO:	Sequence	Fluorescence before phosphorylation (% of wild type)	Relative rates of phosphorylation	%Change in fluorescence after incubation with kinase
19	25RRFSV	95	1.75	-5
20	68RRFSR	0	n.d	n.d
14	68RRFSA	6	0.6	+10
21	94RRSIF	0	n.d	n.d
22	131RRGSIL	0	n.d	n.d
23	155KRKSGI	86	2.5	0
24	172RRGSV	90	1.57	0
18	199RRLSI	0	n.d	n.d
15	214KRDSM	21	1.88	+40

Bold letters indicate site of phosphorylation. Numbers prior to the sequence indicate amino acid position in wild type GFP (Fig. 3, SEQ ID NO:2) where phosphorylation site starts. The relative rates of phosphorylation compare the rate of phosphorylation of the given phosphorylation site with the endogenous protein kinase A phosphorylation site in *Aequorea* GFP (HKFSV SEQ ID NO:1) measured by incorporation of ^{32}P after incubation of the purified substrate and protein kinase A catalytic subunit in the presence of ^{32}P -labelled ATP using 3 μg GFP, 5 μg protein kinase A catalytic subunit for 10 minutes at 30°C in standard phosphorylation buffer (20 mM MOPS pH 6.5, 100mM KCl, 100 μM ATP, 3mM MgCl_2 1 mM DTT and 100uCi ^{32}P -labeled ATP. Reactions were terminated by blotting onto phosphocellulose paper and washing with 10% phosphoric acid. The % change in fluorescence represents the increase in fluorescence (475 nm excitation, 510 nm emission) observed in each purified protein resulting from incubation with excess protein kinase A catalytic subunit for 1 hour at 30° C using the same phosphorylation conditions as described above except that

no ^{32}P -labeled ATP was present and that after the reaction time was complete samples were analyzed in the fluorimeter rather than blotted onto phosphocellulose paper.

The greatest changes in fluorescence occurred in mutant 214KRDSM (SEQ ID NO:15) which exhibited a 40% change in fluorescence upon phosphorylation.

5 However analysis of the kinetics of phosphorylation using γ - ^{32}P -labeled ATP demonstrated that the site is poorly phosphorylated by protein kinase A. Wild type GFP contains a mediocre consensus phosphorylation site (25HKFSV, from SEQ ID NO:1) that can be phosphorylated by protein kinase A *in vitro* with relatively slow kinetics. While phosphorylation at this position has no detectable effect on the fluorescence of GFP, the
10 rate of phosphorylation at this position is used as an internal control between experiments to determine the relative rates of phosphorylation at sites engineered into the protein by site directed mutagenesis.

B. Phosphorylation sites around the amino terminus

15 Sites at the N-terminus of GFP were engineered into GFP by PCR. Initial studies attempted to preserve the native sequence as much as possible. As discussed earlier the positions chosen for phosphorylation were within the first 5 amino acids of GFP and encompassed all charged residues within this region. The sequence changes, relative fluorescence, relative rates of phosphorylation and % change in fluorescence
20 upon phosphorylation are tabulated in Table III.

Table III: Relative fluorescence, rate of phosphorylation and change in fluorescence upon phosphorylation for phosphorylation sites inserted at the N-terminus

25	SEQ ID NO:	Sequence	Relative fluorescence as a % of wild type	Relative rates of phosphorylation	% Change in fluorescence
	2	1MSKGEELF	100	1.0	0
	25	1MRKGSCLF	40	5.1	5.7
30	26	1MRKGSLLF	52	1.6	8.0
	27	1MRRESLLF	30	3.0	6.0
	28	1MRRDSCLF	27	3.7	17
	29	1MSRRDSCF	43	2.1	25
	30	1MSKRRDSL	7	5.5	5.1

Numbers prior to the sequence indicate amino acid position in wild type GFP where phosphorylation site starts. The relative rates of phosphorylation compare the rate of phosphorylation of the given phosphorylation site with the endogenous protein kinase A phosphorylation site in *Aequorea* GFP (HKFSV) measured by incorporation of ³²P after incubation of the purified substrate and protein kinase A catalytic subunit in the presence of ³²P-labelled ATP using the standard protocols described earlier. The % change in fluorescence represents the change in fluorescence (488 nm excitation, 511 nm emission) observed in each purified protein as a result of incubation with excess protein kinase A catalytic subunit for 1 hour at 30° C using phosphorylation conditions described earlier.

These results demonstrated that mutants whose sequence closely resembles the native protein retain considerable fluorescence, display good kinetics of phosphorylation, but show relatively small changes in fluorescence after phosphorylation. To improve the effect of phosphorylation on fluorescence, amino acids around the phosphorylation site were mutated to create an optimal phosphorylation sequence even if it disordered the existing local tertiary structure. Such disruption was predicted and found to decrease the basal fluorescence of these constructs in their non-phosphorylated state (Table IV).

Table IV: Relative fluorescence before phosphorylation and change in fluorescence upon phosphorylation for more drastically altered phosphorylation sites inserted at the N-terminus

SEQ ID NO:	Sequence	Relative fluorescence as a % of wild-type	%Change in fluorescence upon phosphorylation
2	1MSKGEELF (= WT)	= 100	0
31	1MSRRRRSI	5.8	40
32	1MRRRRSII	5.1	70
33	-1MRRRRSIII	n.d.	43
34	-2MRRRRSIIIF	0.7	15
35	-3MRRRRSIIIF	0.6	70

Numbers prior to the sequence indicate amino acid position in wild type GFP where phosphorylation site starts. Negative numbers indicate extensions onto the wild-type N-

terminus. The % change in fluorescence represents the change in fluorescence (488 excitation, 511 emission) observed in each purified protein resulting from incubation with excess protein kinase A catalytic subunit for 1 hour at 30° C using standard phosphorylation conditions described earlier.

5 Perhaps because of the reduced basal fluorescence, phosphorylation by protein kinase A produced greater percentage increases in fluorescence in these constructs than in the more conservative mutations of Table II. Constructs 1MRRRRSII (SEQ ID NO:32) and -3MRRRRSIIIF (SEQ ID NO:35) displayed the greatest increases, about 70%, in fluorescence upon phosphorylation using the standard conditions, as
10 shown in Fig. 6. However, these increased percentage increases were obtained at the cost of a reduced ability to fold at higher temperatures and relatively poor fluorescence even after phosphorylation. To improve these characteristics, these mutants were further optimized by additional random mutagenesis with a novel selection procedure.

15 C. Further optimization of N-terminal phosphorylation sites by random mutagenesis of the remainder of GFP

The two best constructs from above (1MRRRRSII (SEQ ID NO:32) and -3MRRRRSIIIF (SEQ ID NO:35)) were further mutagenized and screened for variants that are highly fluorescent when phosphorylated, but weakly fluorescent when non-
20 phosphorylated. The method involved expression of a randomly mutated fluorescent substrate with or without simultaneous co-expression of the constitutively active catalytic subunit of protein kinase A in bacteria, and screening the individual mutants to determine those that are highly fluorescent in the presence but not the absence of the kinase.

To enable co-expression of the kinase and potential substrates, a new
25 expression vector with the kinase C subunit upstream from the fluorescent substrate was constructed (Fig. 7). Random mutations were introduced into GFP by error-prone PCR and the resulting population of mutants cloned into the co-expression vector using the appropriate restriction sites. The expression vector containing the mutated fluorescent substrates were transformed into host bacteria and individual bacterial colonies (each
30 derived from a single cell, and hence containing a single unique mutant fluorescent substrate) were grown up.

The colonies were screened for fluorescence either by fluorescence-activated cell sorting (Fig. 8) or by observation under a microscope. Those that exhibited the greatest fluorescence were re-screened under conditions in which the kinase gene was inactivated. This was achieved in either of two ways. In the first method the co-expression vector was isolated and treated with restriction endonucleases and modifying enzymes (EcoR1, klenow fragment and T4 DNA ligase) to cut the kinase gene, add additional bases and religate the DNA, causing a frame shift and hence inactivating the gene. The treated and non-treated plasmids were then re-transformed into bacteria and compared in fluorescence. Alternatively the plasmids were initially grown in a RecA⁻ (recombinase A negative) bacterial strain, where the kinase is stable, to screen for brighter mutants in the presence of the kinase. The plasmid DNA was then isolated and re-transformed into a strain of bacteria which is RecA⁺, in which the kinase is unstable and is lost through homologous recombination of the tandemly repeated ribosome binding sites (rbs). The bacteria have a strong tendency to eliminate the kinase C subunit because it slows their multiplication, so cells that splice out the kinase by recombination have a large growth advantage.

Comparison of the brightness of the mutant first in the presence of kinase then in its absence indicates the relative effect of phosphorylation on the mutant GFP fluorescence (after normalizing for GFP expression levels). A library of approximately 2 x10⁶ members was screened by this approach. Approximately 500 displayed higher levels of fluorescence when screened in the presence of the kinase. After inactivation of the kinase, one mutant out of the 500 displayed reduced levels of fluorescence. The increased fluorescence of the remainder of the 500 mutants was independent of the presence of the kinase. This mutant GFP was isolated and sequenced and found to contain the following mutations compared to wild-type GFP (Fig. 3, SEQ ID NO:2) (in addition to the N-terminal phosphorylation site 1MRRRRSII (SEQ ID NO:32)): S65A, N149K, V163A and I167T.

To confirm that this mutant was indeed directly sensitive to protein kinase A phosphorylation and to quantify its responsiveness, it was expressed in the absence of kinase. The *E. coli* were lysed and the protein purified as described earlier using a nickel affinity column. The protein exhibited high levels of fluorescence when induced at 30° C but displayed reduced fluorescence when incubated at 37°C. After such preincubation (37° C overnight) and separation of the less fluorescent material by

centrifugation, this protein exhibited the largest change in fluorescence upon phosphorylation yet observed (Fig. 8). The tolerance of this mutant for 37°C treatment suggests that this mutant is suitable for use in mammalian cells.

5 The present invention provides novel assays for protein kinase activity involving novel fluorescent protein substrates. While specific examples have been provided, the above description is illustrative and not restrictive. Many variations of the invention will become apparent to those skilled in the art upon review of this specification. The scope of the invention should, therefore, be determined not with
10 reference to the above description, but instead should be determined with reference to the appended claims along with their full scope of equivalents.

 All publications and patent documents cited in this application are incorporated by reference in their entirety for all purposes to the same extent as if each individual publication or patent document were so individually denoted.

15

WHAT IS CLAIMED IS:

- 2 1. A method for determining whether a sample contains protein kinase
3 activity comprising:
4 contacting the sample with a phosphate donor and a fluorescent
5 protein substrate for a protein kinase, the protein substrate comprising a fluorescent
6 protein moiety and a phosphorylation site for a protein kinase, wherein the protein
7 substrate exhibits a different fluorescent property in the phosphorylated state than in the
8 un-phosphorylated state;
9 exciting the protein substrate; and
10 measuring the amount of a fluorescent property that differs in the
11 un-phosphorylated state and phosphorylated state, whereby an amount that is consistent
12 with the presence of the protein substrate in its phosphorylated state indicates the
13 presence of protein kinase activity.
- 1 2. The method of claim 1 for determining the amount of protein
2 kinase activity in a sample wherein measuring the amount of a fluorescent property in the
3 sample comprises measuring the amount at two or more time points after contacting the
4 sample with a phosphate donor and a fluorescent protein substrate, and determining the
5 quantity of change or rate of change of the measured amount, whereby the quantity or
6 rate of change of the measured amount reflects the amount of protein kinase activity in
 the sample.
- 1 3. The method of claim 2 wherein the fluorescent property is the
2 fluorescent emission around the emission maximum of the substrate in the phosphorylated
 state.
- 1 4. The method of claim 2 wherein amount is determined by emission
2 amplitude ratioing or excitation amplitude ratioing.
- 1 5. The method of claim 3 wherein the fluorescent protein is an
2 *Aequorea*-related fluorescent protein.

1 6. A method for determining whether a cell exhibits protein kinase
2 activity comprising the steps of:
3 providing a transfected host cell comprising a recombinant nucleic
4 acid molecule comprising expression control sequences operatively linked to a nucleic
5 acid sequence coding for the expression of a fluorescent protein substrate for a protein
6 kinase, the protein substrate comprising a fluorescent protein moiety containing a
7 phosphorylation site for a protein kinase, wherein the protein substrate exhibits a
8 different fluorescent property in the phosphorylated state than in the un-phosphorylated
9 state, the cell expressing the fluorescent protein substrate;
10 exciting the protein substrate in the cell; and
11 measuring the amount of a fluorescent property that differs in the
12 un-phosphorylated and phosphorylated states, wherein the presence of the fluorescent
13 property associated with the fluorescent state indicates the presence of protein kinase
14 activity in the cell.

1 7. The method of claim 6 wherein the fluorescent property is the
2 fluorescent emission around the emission maximum of the substrate in the phosphorylated
 state.

1 8. The method of claim 6 wherein the amount is determined by
2 emission amplitude ratioing or excitation amplitude ratioing.

1 9. The method of claim 6 wherein the cell is further transfected with
2 an expression vector comprising expression control sequences operatively linked to a
3 nucleic acid sequence coding for the expression of the protein kinase.

1 10. The method of claim 6 wherein the fluorescent protein is an
2 *Aequorea*-related fluorescent protein.

1 11. The method of claim 6 wherein the step of providing a transfected
2 host cell comprises inducing expression of the protein substrate to produce a sudden
3 increase in the expression of the protein substrate, and the step of measuring the amount
4 of a fluorescent property comprises measuring the amount at a first and a second time
5 after expression of the protein substrate and determining the difference between the
6 measured amounts at the first and second time.

1 12. A method for determining the amount of activity of a protein kinase
2 in a sample from an organism comprising the steps of:
3 providing a sample from an organism having a cell that expresses a
4 fluorescent protein substrate for a protein kinase, the protein substrate comprising a
5 fluorescent protein moiety and a phosphorylation site for a protein kinase, wherein the
6 protein substrate exhibits a different fluorescent property in the phosphorylated state than
7 in the un-phosphorylated state;
8 contacting the sample with a phosphate donor;
9 exciting the protein substrate; and
10 measuring the amount of a fluorescent property that differs in the
11 un-phosphorylated state and phosphorylated state, whereby an amount that is consistent
12 with the presence of the protein substrate in its phosphorylated state indicates the
13 presence of protein kinase activity, and an amount that is consistent with the presence of
14 the protein substrate in its un-phosphorylated state indicates the absence of protein kinase
15 activity.

1 13. The method of claim 12 wherein the amount is determined by
2 emission amplitude ratioing or excitation amplitude ratioing.

1 14. The method of claim 12 wherein the fluorescent protein is an
2 *Aequorea*-related fluorescent protein.

1 15. The method of claim 12 wherein the sample is a cell homogenate.

1 16. A method for determining whether a compound alters the activity
2 of a protein kinase comprising the steps of:

3 contacting a sample containing a known amount of protein kinase
4 activity with the compound, a phosphate donor for the protein kinase and a fluorescent
5 protein substrate for a protein kinase, the protein substrate comprising a fluorescent
6 protein moiety and a phosphorylation site for a protein kinase, wherein the protein
7 substrate exhibits a different fluorescent property in the phosphorylated state than in the
8 un-phosphorylated state;

9 exciting the protein substrate;

10 measuring the amount of protein kinase activity in the sample as a
11 function of the quantity of change or rate of change of a fluorescent property that differs
12 in the un-phosphorylated and phosphorylated states; and

13 comparing the amount of activity in the sample with a standard
14 activity for the same amount of the protein kinase, whereby a difference between the
15 amount of protein kinase activity in the sample and the standard activity indicates that the
16 compound alters the activity of the protein kinase.

1 17. The method of claim 16 wherein the amount is determined by
2 emission amplitude ratioing or excitation amplitude ratioing.

1 18. The method of claim 16 wherein the fluorescent protein is an
2 *Aequorea*-related fluorescent protein.

1 19. A method for determining whether a compound alters the protein
2 kinase activity in a cell comprising the steps of:

3 providing first and second transfected host cells exhibiting protein
4 kinase activity and expressing a fluorescent protein substrate for a protein kinase, the
5 protein substrate comprising a fluorescent protein moiety and a phosphorylation site for a
6 protein kinase, wherein the protein substrate exhibits a different fluorescent property in
7 the phosphorylated state than in the un-phosphorylated state;

8 contacting the first cell with an amount of the compound;

9 contacting the second cell with a different amount of the compound;
10 exciting the protein substrate in the first and second cells;
11 measuring the amount of protein kinase activity in the cells as a
12 function of the quantity of change or rate of change of a fluorescent property that differs
13 in the un-phosphorylated and phosphorylated states in the first and second cells; and
14 comparing the amount in the first and second cells, whereby a
15 difference in the amount indicates that the compound alters protein kinase activity in the
16 cell.

1 20. The method of claim 19 wherein the amount is determined by
2 emission amplitude ratioing or excitation amplitude ratioing.

1 21. The method of claim 19 wherein the fluorescent protein is an
2 *Aequorea*-related fluorescent protein.

1 22. The method of claim 18 wherein the cells are transfected with an
2 expression vector comprising expression control sequences operatively linked to a nucleic
3 acid sequence coding for the expression of the protein kinase.

1 23. A fluorescent protein substrate for a protein kinase comprising a
2 fluorescent protein moiety and a phosphorylation site for a protein kinase, wherein the
3 protein substrate exhibits a different fluorescent property in the phosphorylated state than
4 in the un-phosphorylated state.

1 24. The protein substrate of claim 23 wherein the fluorescent protein is
2 an *Aequorea*-related fluorescent protein.

1 25. The protein substrate of claim 24 comprising a phosphorylation site
2 for protein kinase A, a cGMP-dependent protein kinase, protein kinase C,
3 Ca²⁺/calmodulin-dependent protein kinase I, Ca²⁺ / calmodulin-dependent protein kinase
4 II or MAP kinase activated protein kinase type 1.

1 26. The protein substrate of claim 25 wherein the phosphorylation site
2 is RRXSZ (SEQ ID NO:3) or RRXTZ (SEQ ID NO:4), wherein X is any amino acid
3 and Z is a hydrophobic amino acid, BKISASEFDR PLR (SEQ ID NO:5), where B
4 represents either lysine (K) or arginine (R), and the first S is the site of phosphorylation,
5 XRXXSXRX (SEQ ID NO:7), wherein X is any amino acid, KKKKRFSFK (SEQ ID
6 NO:8), LRRLSDSNF (SEQ ID NO:9), KKLNRSLTVA (SEQ ID NO:10),
7 KKANRTLVA (SEQ ID NO:11).

1 27. The protein substrate of claim 19 wherein the *Aequorea*-related
2 fluorescent protein is variant P4, P4-3, W7, W2, S65T, P4-1, S65A, S65L, Y66F or
3 Y66W.

1 28. The protein substrate of claim 24 wherein the *Aequorea*-related
2 fluorescent protein comprises a folding mutation.

1 29. The protein substrate of claim 24 comprising the phosphorylation
2 site within twenty amino acids of the amino-terminus of the fluorescent protein moiety.

1 30. The protein substrate of claim 23 wherein the phosphorylation site
2 is contained within the sequence MRRRRSIITG (SEQ ID NO:12) or MRRRRSI IFTG
3 (SEQ ID NO:13).

1 31. The protein substrate of claim 30 further comprising the
2 substitutions S65A, N149K, V163A and I167T.

1 32. The protein substrate of claim 24 comprising the phosphorylation
2 site within the fluorescent protein moiety more than twenty amino acids from the amino
 terminus.

- 1 33. The protein substrate of claim 32 comprising the substitution
2 H217S.
- 1 34. The protein substrate of claim 32 wherein the sequence of wild type
2 *Aequorea* GFP is modified by the substitution 69RRFSA (SEQ ID NO:14) or
3 214KRDSM (SEQ ID NO:15).
- 1 35. The protein substrate of claim 32 further comprising the
2 substitution E171K and/or I172V.
- 1 36. The protein substrate of claim 24 wherein the protein substrate is a
2 fusion protein.
- 1 37. A nucleic acid molecule coding for the expression of a fluorescent
2 protein substrate for a protein kinase comprising a fluorescent protein moiety and a
3 phosphorylation site for a protein kinase, wherein the protein substrate exhibits a
4 different fluorescent property in the phosphorylated state than in the un-phosphorylated
5 state.
- 1 38. The nucleic acid molecule of claim 37 wherein the fluorescent
2 protein is an *Aequorea*-related fluorescent protein.
- 1 39. The nucleic acid molecule of claim 37 wherein the protein substrate
2 comprises a phosphorylation site for protein kinase A, a cGMP-dependent protein kinase,
3 protein kinase C, Ca²⁺/calmodulin-dependent protein kinase I, Ca²⁺/ calmodulin-
4 dependent protein kinase II or MAP kinase activated protein kinase type 1.
- 1 40. The nucleic acid molecule of claim 37 wherein the phosphorylation
2 site is RRXSZ (SEQ ID NO:3) or RRXTZ (SEQ ID NO:4), wherein X is any amino acid
3 and Z is a hydrophobic amino acid, BKISASEFDR PLR (SEQ ID NO:5), where B
4 represents either lysine (K) or arginine (R), and the first S is the site of phosphorylation,

5 XRXXSXRX (SEQ ID NO:7), wherein X is any amino acid, KKKKRFSFK (SEQ ID
6 NO:8), LRRLSDSNF (SEQ ID NO:9), KKLNRSLTVA (SEQ ID NO:10),
7 KKANRTLSVA (SEQ ID NO:11).

1 41. The nucleic acid molecule of claim 37 wherein the *Aequorea*-
2 related fluorescent protein is variant P4, P4-3, W7, W2, S65T, P4-1, S65A, S65L,
3 Y66F or Y66W.

1 42. The nucleic acid molecule of claim 37 wherein the *Aequorea*-
2 related fluorescent protein comprises a folding mutation.

1 43. The nucleic acid molecule of claim 37 wherein the protein substrate
2 comprises the phosphorylation site within twenty amino acids of the amino-terminus of
3 the fluorescent protein moiety.

1 44. The nucleic acid molecule of claim 37 wherein the phosphorylation
2 site is contained within the sequence MRRRRSIITG (SEQ ID NO:12) or MRRRRSII
3 IIFTG (SEQ ID NO:13).

1 45. The nucleic acid molecule of claim 37 wherein the protein substrate
2 further comprises the substitutions S65A, N149K, V163A and I167T.

1 46. The nucleic acid molecule of claim 37 wherein the protein substrate
2 comprises the phosphorylation site within the fluorescent protein moiety more than
 twenty amino acids from the amino terminus.

1 47. The nucleic acid molecule of claim 37 wherein the protein substrate
2 comprises the substitution H217S.

1 48. The nucleic acid molecule of claim 37 wherein the sequence of
2 wild type *Aequorea* GFP is modified by the substitution 69RRFSA (SEQ ID NO:14) or
3 214KRDSM (SEQ ID NO:15).

1 49. The nucleic acid molecule of claim 37 wherein the protein substrate
2 further comprises the substitution E171K and/or I172V.

1 50. A recombinant nucleic acid molecule comprising expression control
2 sequences operatively linked to a nucleic acid sequence coding for the expression of a
3 fluorescent protein substrate for a protein kinase comprising a fluorescent protein moiety
4 and a phosphorylation site for a protein kinase, wherein the protein substrate exhibits a
5 different fluorescent property in the phosphorylated state than in the un-phosphorylated
6 state.

1 51. A transfected host cell comprising a recombinant nucleic acid
2 molecule comprising expression control sequences operatively linked to a nucleic acid
3 sequence coding for the expression of a fluorescent protein substrate for a protein kinase
4 comprising a fluorescent protein moiety and a phosphorylation site for a protein kinase,
5 wherein the protein substrate exhibits a different fluorescent property in the
6 phosphorylated state than in the un-phosphorylated state.

1 52. A kit comprising a fluorescent protein substrate for a protein kinase
2 comprising a fluorescent protein moiety and a phosphorylation site for a protein kinase,
3 and comprising a phosphate donor.

1 53. A collection of fluorescent protein candidate substrates comprising
2 at least 10 different members, each member comprising a fluorescent protein moiety and
3 a variable peptide moiety within about twenty amino acids of the amino-terminus of the
4 fluorescent protein moiety.

1 54. The collection of claim 53 containing at least 10^3 different
2 members.

1 55. The collection of claim 54 containing at least 10^6 different
2 members.

1 56. A collection of recombinant nucleic acid molecules comprising at
2 least 10 different recombinant nucleic acid molecule members, each member comprising
3 expression control sequences operatively linked to nucleic acid sequences coding for the
4 expression of a different fluorescent protein candidate substrate which comprises a
5 fluorescent protein moiety and a variable peptide moiety within about twenty amino acids
6 of the amino-terminus of the fluorescent protein moiety.

1 57. A collection of host cells comprising at least 10 different host cell
2 members, each member comprising a recombinant nucleic acid molecule which
3 comprises expression control sequences operatively linked to nucleic acid sequences that
4 code for the expression of a different fluorescent protein candidate substrate which
5 comprises a fluorescent protein moiety and a variable peptide moiety within about twenty
6 amino acids of the amino-terminus of the fluorescent protein moiety.

1 58. A method for screening a collection of transfected host cells
2 comprising:
3 providing a collection of transfected host cells comprising at least
4 10 different host cell members, each member expressing a different fluorescent protein
5 candidate substrate, the candidate substrate comprising a fluorescent protein moiety and a
6 variable peptide moiety within about twenty amino acids of the amino-terminus of the
7 fluorescent protein moiety, wherein the fluorescent protein exhibits a fluorescent
8 property;
9 measuring the fluorescent property in a sample comprising at least
10 one host cell member before and after increasing or decreasing the intracellular protein
11 kinase activity in the cell; and

12 determining the degree of change or the rate of change in the
13 fluorescent property upon increasing or decreasing intracellular protein kinase activity;
14 whereby a change in the degree or rate of the fluorescent property
15 indicates that the candidate substrate possesses a peptide moiety that can be
16 phosphorylated by the protein kinase and whose phosphorylation alters the fluorescent
17 property.

1 59. The method of claim 58 whereby the terminus is the amino-
2 terminus.

3 60. The method of claim 59 further comprising determining the change
4 in a plurality of host cell members and comparing the degree of change or rate of change
5 between the host cell members; whereby the comparison indicates in the candidate
6 substrates the relative change in the fluorescent property upon phosphorylation.

1 61. The method of claim 60 further comprising isolating a member
2 having the altered fluorescent property.

1 62. The method of claim 59 wherein the step of determining the rate of
2 change comprises measuring the fluorescent property at a plurality of time points after
3 inducing intracellular protein kinase activity.

1 63. The method of claim 59 wherein sample comprises a clonal
2 expansion of the host cell.

1 64. The method of claim 59 wherein the host cell is co-transfected with
2 an expression vector that expresses a protein kinase comprising expression control
3 sequences operatively linked to a sequence that codes for the expression of the protein
4 kinase.

1 65. The method of claim 64 wherein the step of increasing the
2 intracellular protein kinase activity comprises elevating intracellular cAMP to activate
3 cAMP-dependent protein kinase, elevating intracellular cGMP to activate cGMP-
4 dependent protein kinase, elevating cytosolic free calcium to activate Ca²⁺/calmodulin-
5 dependent protein kinase types I, II, or IV, or administration of phorbol myristate acetate
6 to activate protein kinase C.

1 66. The method of claim 59 wherein the step of decreasing the
2 intracellular protein kinase activity comprises culturing the cell for a time sufficient for
3 the cell to lose sequence that codes for the expression of the protein kinase.

1 67. The method of claim 59 further comprising isolating the
2 recombinant nucleic acid molecule coding for the expression of the candidate substrate.

1 68. A method for screening a collection of transfected host cells
2 comprising:
3 providing a lysate from each of at least one member of a collection
4 of transfected host cells comprising at least 10 different host cell members, each member
5 expressing a different fluorescent protein candidate substrate, the candidate substrate
6 comprising a fluorescent protein moiety and a variable peptide moiety around the amino-
7 terminus of the fluorescent protein moiety, wherein the fluorescent protein exhibits a
8 fluorescent property;
9 contacting the lysate with a phosphate donor;
10 measuring the fluorescent property from at least one lysate before
11 and after contacting the lysate with a protein kinase; and
12 determining the degree of change or the rate of change in the
13 fluorescent property upon contacting the lysate with intracellular protein kinase activity;
14 whereby a change in the degree or rate of the fluorescent property
15 indicates that the candidate substrate possesses a peptide moiety that can be
16 phosphorylated by the protein kinase and whose phosphorylation alters the fluorescent
17 property.

1 69. The method of claim 68 comprising splitting the lysate into first
2 and second aliquots;
3 contacting the first and second aliquot with a phosphate donor;
4 contacting the first aliquot with a protein kinase but not contacting
5 the second aliquot with the protein kinase;
6 measuring the fluorescent property from the first aliquot before and
7 after contacting the lysate with the protein kinase;
8 measuring the fluorescent property from the second aliquot at a
9 plurality of time points after contact with the phosphate donor;
10 determining the degree of change or the rate of change in the
11 fluorescent property of the first and second aliquots;
12 comparing the degree or rate of change in the first and second
13 aliquots;
14 whereby a difference in the degree or rate of change in the
15 fluorescent property in first and second aliquots indicates that the candidate substrate
16 possesses a peptide moiety that alters the fluorescent property.

1 70. The method of claim 68, wherein the amount is determined by
 emission amplitude ratioing or excitation amplitude ratioing.

1 71. The method of claim 69 wherein the step of determining the rate of
2 change comprises measuring the fluorescent property at a plurality of time points after
3 inducing intracellular protein kinase activity.

1 72. The method of claim 69 wherein the host cell is expanded into a
2 clonal population.

1 73. The method of claim 69 further comprising isolating a member of
2 the library having an altered fluorescent property.

1 74. The method of claim 72 further comprising isolating the
recombinant nucleic acid molecule coding for the expression of the candidate substrate.

1 75. A method of introducing a phosphorylation site into a fluorescent
2 protein comprising the step of genetically attaching an amino acid sequence including a
3 phosphorylation site within twenty amino acids of a terminus of a fluorescent protein
moiety.

1 76. The method of claim 75 wherein the terminus is the amino-
2 terminus.

FIG 1

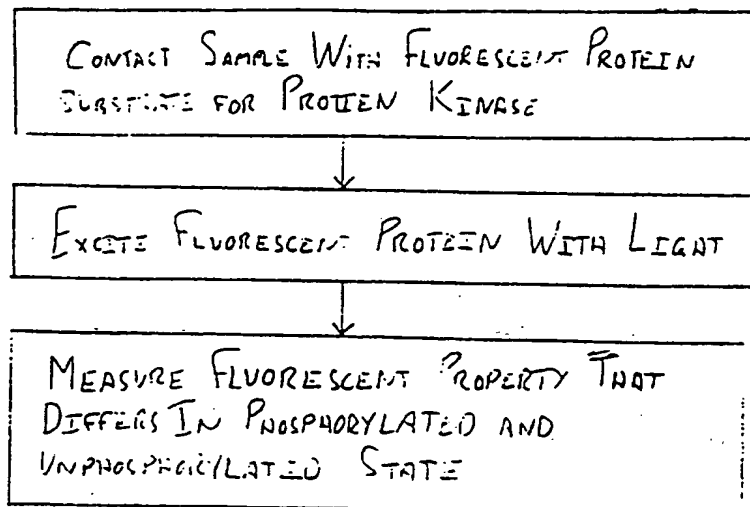
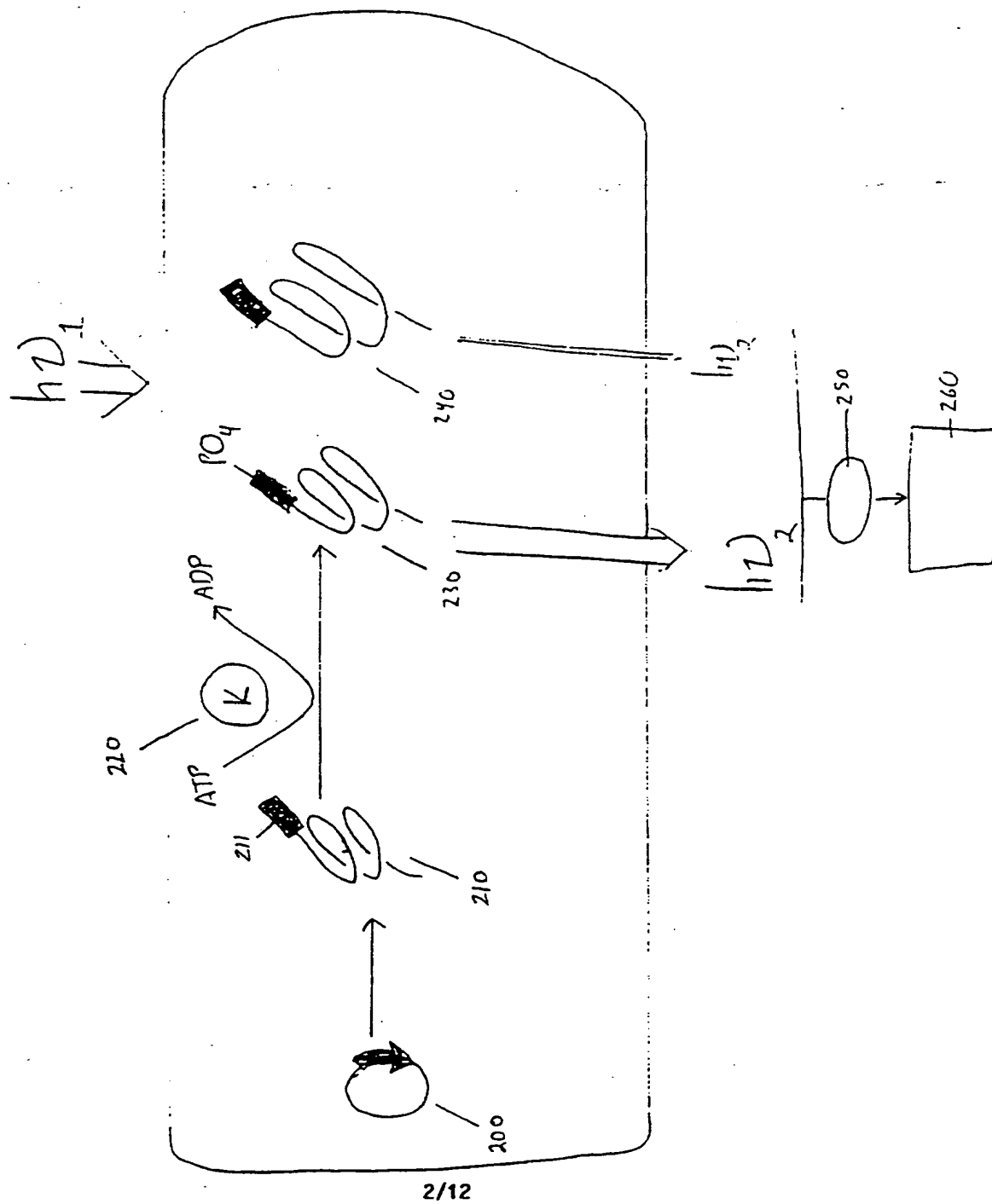


FIG 2



(x1) SEQUENCE DESCRIPTION:

SEQ ID NO:1:	ATG AGT AAA GGA GAA GAA CTT TTC ACT GGA GTT GTC CCA ATT CTT GTT	48
SEQ ID NO:2:	Met Ser Lys Gly Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val	
	1 5 10 15	
	GAA TTA CAT GGT CAT GTT AAT GGG CAC AAA TTT TCT CTC AGT GGA GAG	96
	Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu	
	20 25 30	
	GGT GAA GGT CAT GCA ACA TAC GGA AAA CTT ACC CTT AAA TTT ATT TGC	144
	Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys	
	35 40 45	
	ACT ACT GGA AAA CTA CCT GTT CCA TGG CCA ACA CTT CTC ACT ACT TTC	192
	Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Phe	
	50 55 60	
	TCT TAT GGT GTT CAA TGC TTT TCA AGA TAC CCA GAT CAT ATG AAA GGG	240
	Ser Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Arg	
	65 70 75 80	
	CAT GAC TTT TTC AAG AGT GGC ATG CCC GAA GGT TAT GTA CAG GAA ACA	288
	His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg	
	85 90 95	
	ACT ATA TTT TTC AAA CAT GAC GGG AAC TAC AAG ACA CGT CCT GAA GTC	336
	Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val	
	100 105 110	
	AAG TTT GAA GGT CAT ACC CTT GTT AAT AGA ATC GAG TTA AAA GGT ATT	384
	Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile	
	115 120 125	
	GAT TTT AAA GAA CAT GGA AAC ATT CTT GCA CAC AAA TTC GAA TAC AAC	432
	Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn	
	130 135 140	
	TAT AAC TCA CAC AAT GTA TAC ATC ATG GGA GAC AAA CAA AAG AAT GGA	480
	Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly	
	145 150 155 160	
	ATC AAA GTT AAC TTC AAA ATT AGA CAC AAC ATT GAA GAT GGA AGC GTT	528
	Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val	
	165 170 175	
	CAA CTA GGA GAC CAT TAT CAA CAA AAT ACT CCA ATT GCG CAT GCG CCT	576
	Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro	
	180 185 190	
	GTC CTT TTA CCA GAC AAC CAT TAC CTG TCC ACA CAA TCT GCC CTT TCC	624
	Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser	
	195 200 205	
	AAA GAT CCC AAC GAA AAG AGA GAC CAC ATG GTC CTT CTT CAG TTT GTA	672
	Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val	
	210 215 220	
	ACA GCT GCT GGG ATT ACA CAT GGC ATG CAT GAA CTA TAC AAA TA	717
	Thr Ala Ala Gly Ile Thr His Gly Met Asp Glu Leu Tyr Lys	
	225 230 235	

FIGURE 3

Fig. 4

4

Figure 2. Locations of phosphorylation sites distal to the N-terminus. Amino acids underlined represent the phosphorylation motif, amino acids in brackets represent wild type sequence at those positions.

```

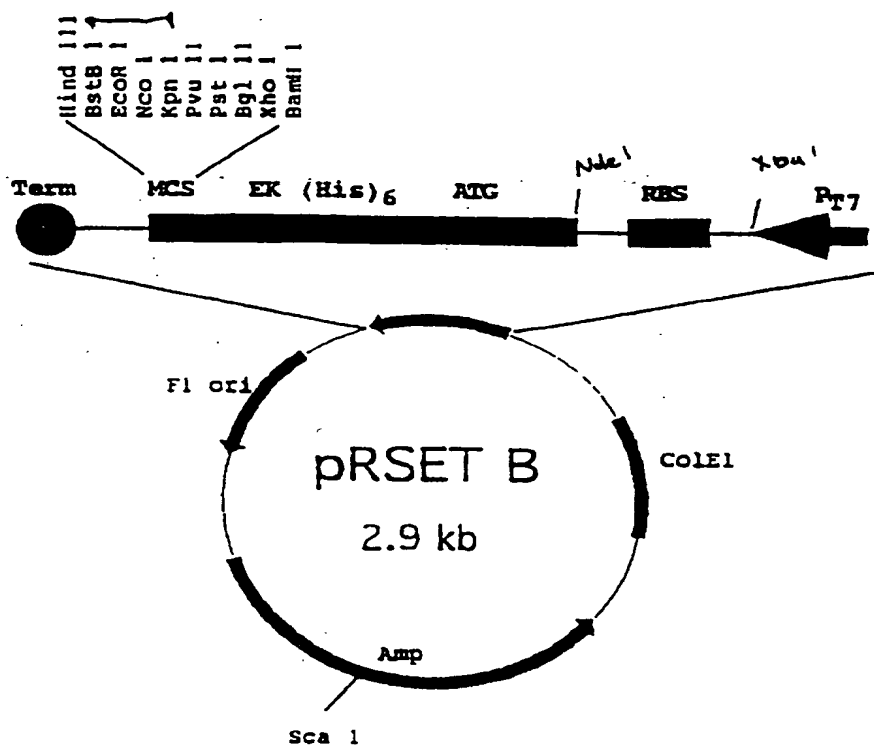
      20              40              60
      .              .              .
ATG AGT AAA GGA GAA GAA CTT TTC ACT CGA GTT GTC CCA ATT CTT GTT GAA TTA GAT GGT
Met Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly
      80              100              120
      (His Lys) .
GAT GTT AAT GGG AGA AGA TTT TCT GTC AGT GGA GAG GGT CAA CGT GAT GCA ACA TAC GGA
Asp Val Asn Gly Arg Arg Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly
      140              160              180
      .
AAA CTT ACC CTT AAA TTT ATT TGC ACT ACT GGA AAA CTA CCT GTT CCA TGG CCA ACA CTT
Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu
      200              220              240
      .
GTC ACT ACT TTC TCT TAT GGT GTT AGA AGA TTT TCA GCA TAC CCA GAT CAT ATG AAA CAG
Val Thr Thr Phe Ser Tyr Gly Val Arg Arg Phe Ser Ala Tyr Pro Asp His Met Lys Gln
      260              280              300
      .
CAT GAC TTT TTC AAG AGT GCC ATG CCC GAA GGT TAT GTA CAG AGA AGA TCT ATA TTT TTC
His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Arg Arg Ser Ile Phe Phe
      320              340              360
      .
AAA GAT GAC GGG AAC TAC AAG ACA CGT GCT GAA GTC AAG TTT GAA GST GAT ACC CTT GTT
Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val
      380              400              420
      .
AAT AGA ATC GAG TTA AAA GGT ATT GAT TTT AAA AGA AGA GGA AAC ATT CTT GGA CAC AAA
Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Arg Arg Gly Ser Ile Leu Gly His Lys
      440              460              480
      .
TTG GAA TAC AAC TAT AAC TCA CAC AAT GTA TAC ATC ATG GCA GAC AAA AGA AAG TCT GGA
Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Arg Lys Ser Gly
      500              520              540
      .
ATC AAA GTT AAC TTC AAA ATT AGA CAC AAC ATT AGA AGA GGA AGC GTT CAA CTA GCA GAC
Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Arg Arg Gly Ser Val Gln Leu Ala Asp
      560              580              600
      .
CAT TAT CAA CAA AAT ACT CCA ATT GGC GAT GGC CCT GTC CTT TTA CCA GAC AAC AGA AGA
His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn Arg Arg
      620              640              660
      .
CTG TCC ATA CAA TCT GCC CTT TCG AAA GAT CCC AAC GAA AAG AGA GAC AGA ATG GTC CTT
Leu Ser Ile Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp Ser Met Val Leu
      680              700
      .
CTT GAG TTT GTA ACA GCT GCT GGG ATT ACA CAT GGC ATG GAT GAA CTA TAC AAA TAA
Leu Glu Phe Val Thr Ala Ala Gly Ile Thr His Gly Met Asp Glu Leu Tyr Lys ***

```

Fig. 5

GFP Bacteria Expression Cassette

The BamHI fragment of GFP in Bluescript 11 was cloned into the BamHI site of pRSET_B (from Invitrogen)



NdeI ProBond Binding Domain XbaI
 GGAGATATACAT ATG CCG GGT TCT CAT CAT CAT CAT CAT CAT GGT ATG GCT AGC ATG ACT GGT GGA CAG
 Met Arg Gly Ser His His His His His His Gly Met Ala Ser Met Thr Gly Gly Gln

 EK Cleavage Site BamHI EcoRI GFP →
 CAA ATG GGT CCG CAT CTC TAC GAC GAT GAC GAT AAG GAT CCC CCC GCT GAA TTC ATG AGT.....TAC
 Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp Pro Pro Ala Glu Phe Met Ser.....Tyr

 BamHI SacI XhoI BglII PstI PvuII KpnI MspI EcoRI BstBI HindIII
 AAA TAA TAA CGATCCGACCTCGAGATCTGCAGCTGGTACCATGGAATTCGAAGGTTGA
 Lys . . .

Fig. 6A

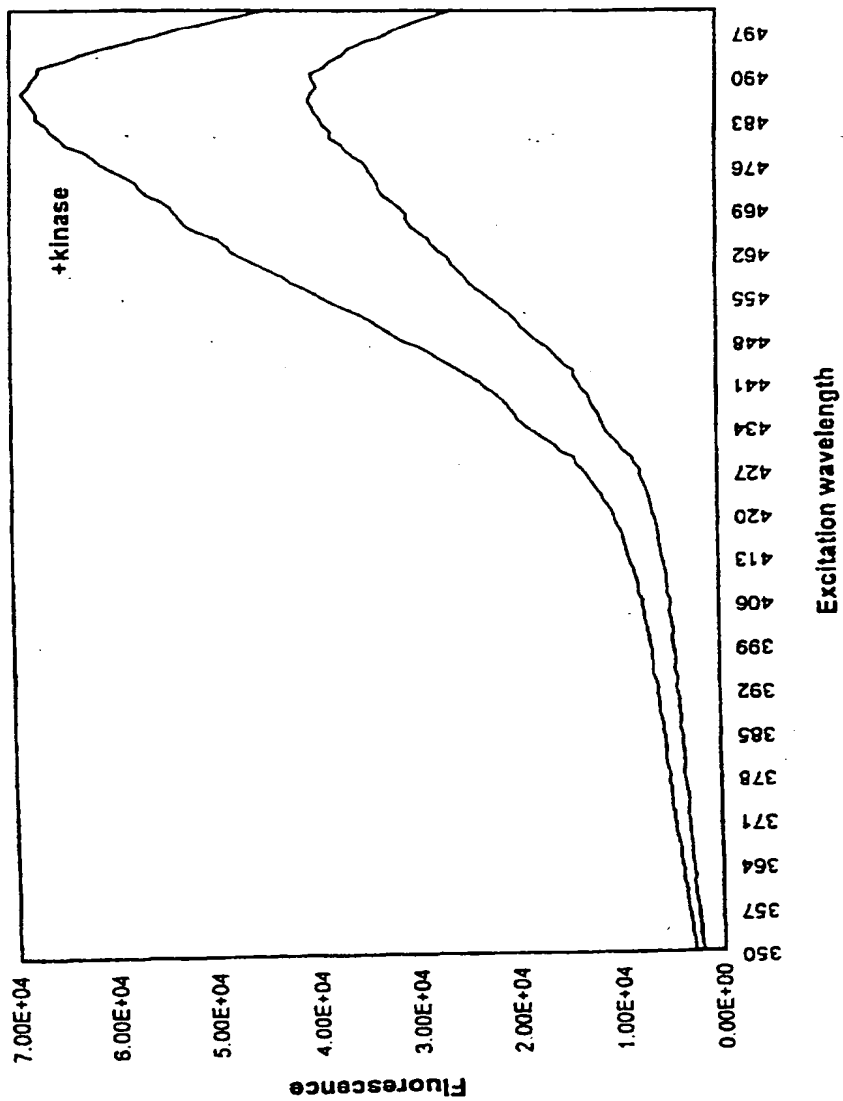


Fig. 6B

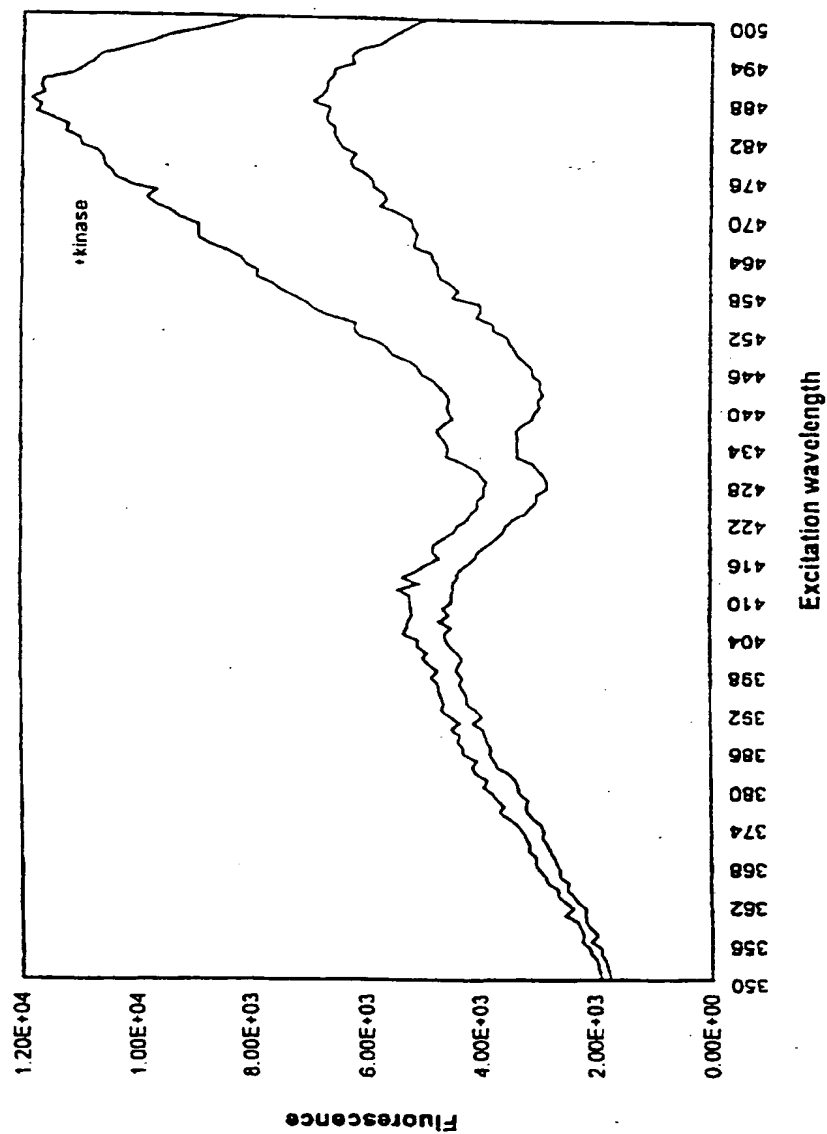


Fig. 6C

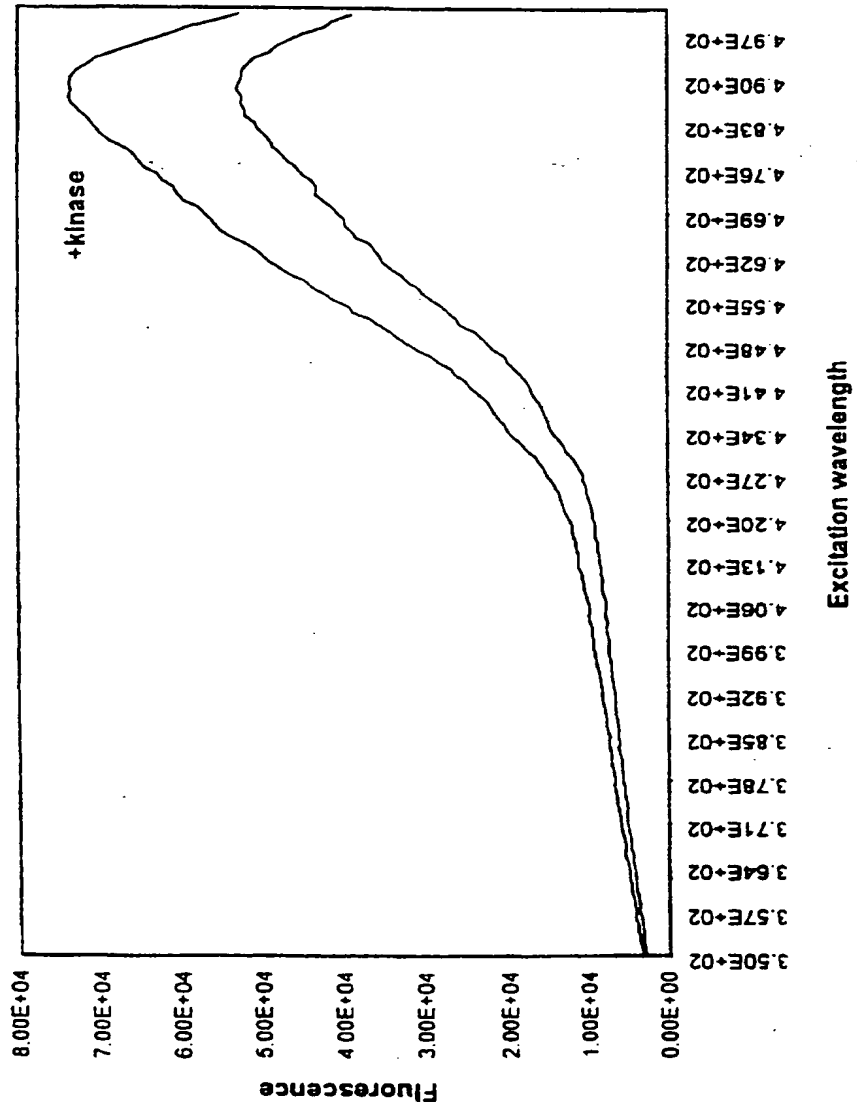


Fig. 6D

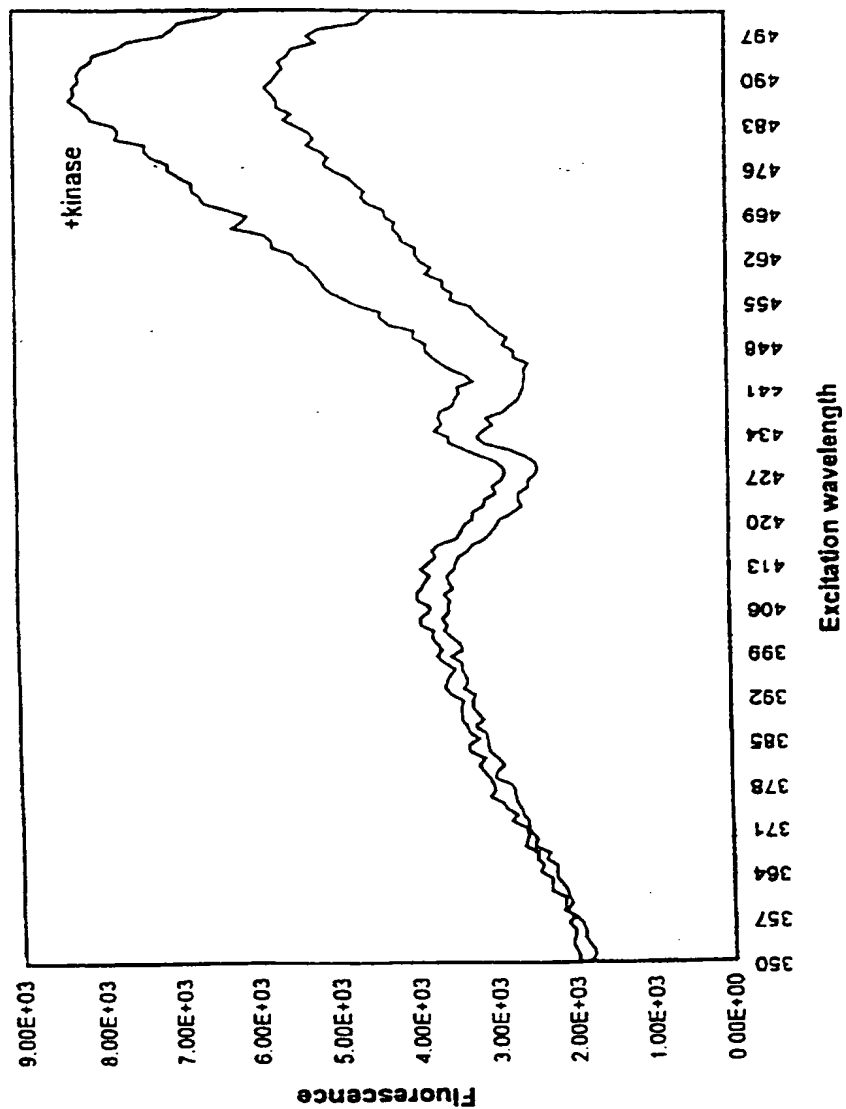


Fig. 6E

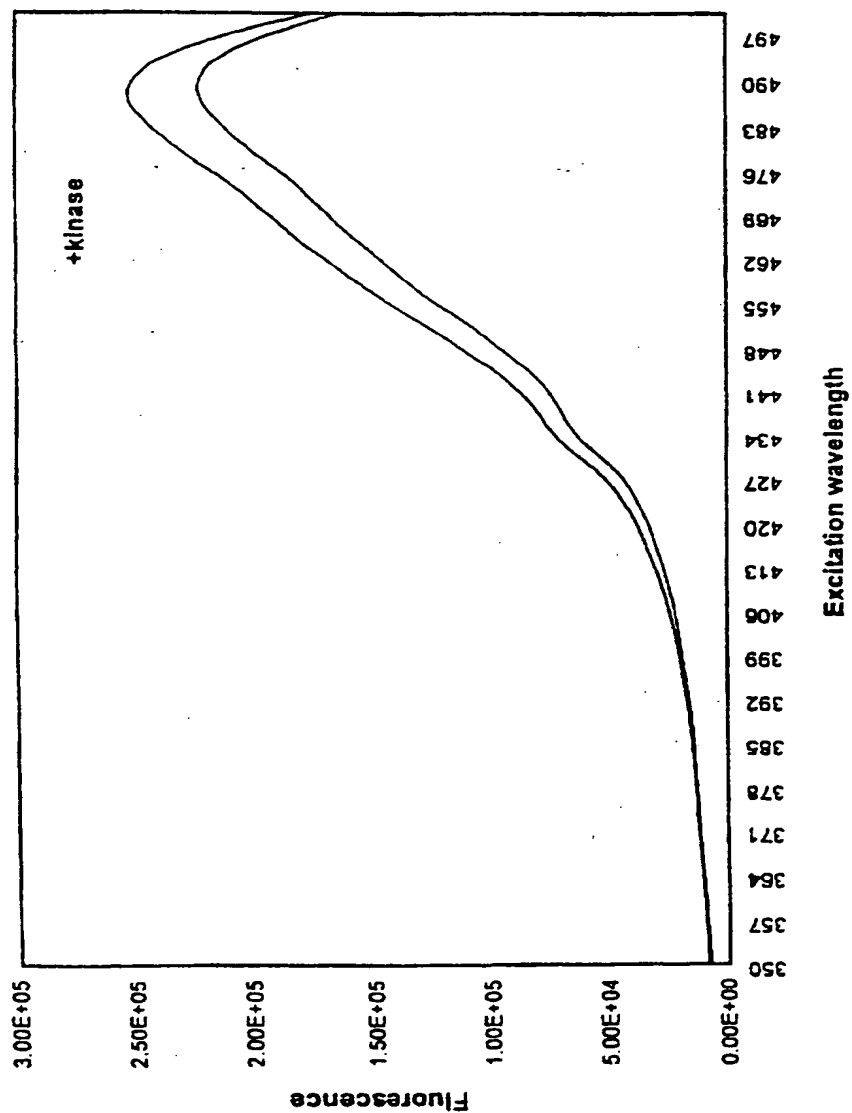
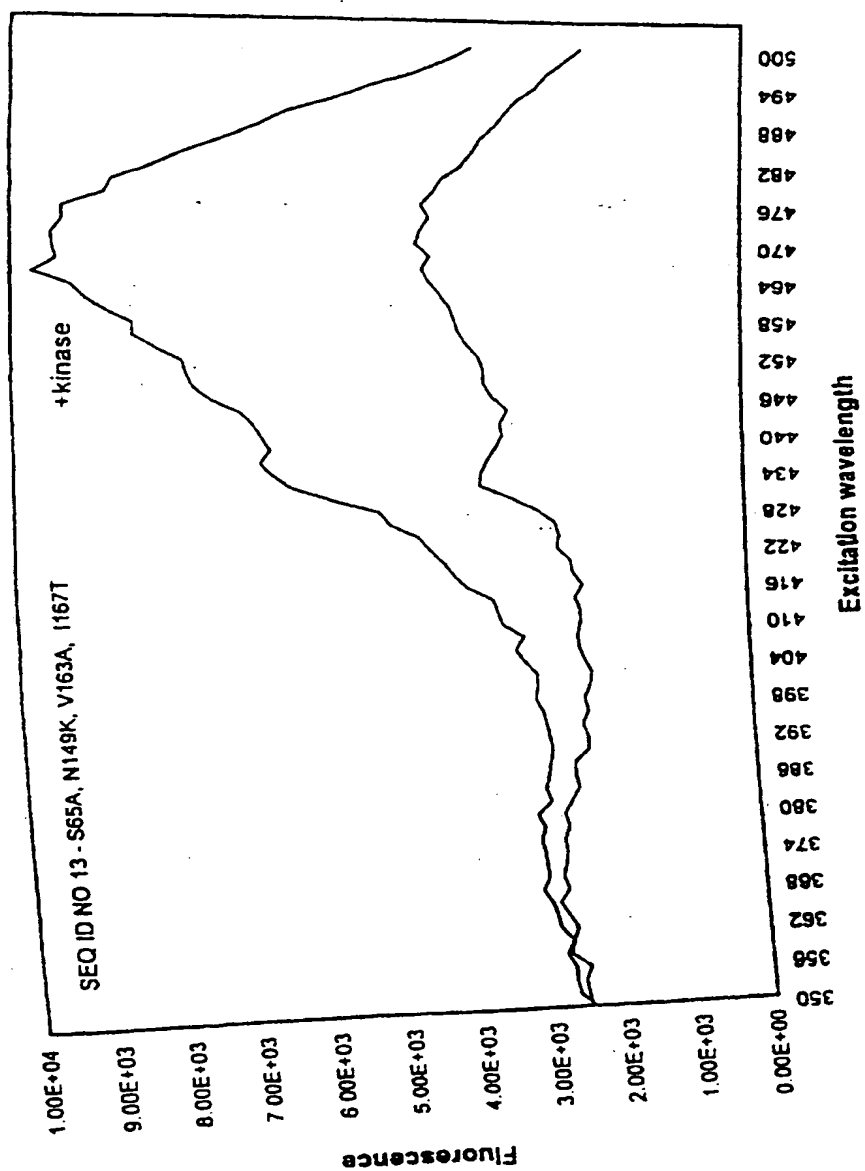


Fig. 8



INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/12410

A. CLASSIFICATION OF SUBJECT MATTER IPC(6) : Please See Extra Sheet. US CL : 536/23.1, 23.4; 435/252.3, 320.1, 172.3, 6, 15; 530/350, 352 According to International Patent Classification (IPC) or to both national classification and IPC																									
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 536/23.1, 23.4; 435/252.3, 320.1, 172.3, 6, 15; 530/350, 352 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Extra Sheet.																									
C. DOCUMENTS CONSIDERED TO BE RELEVANT																									
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.																							
X, P — Y, P	WO 96/23898 A1 (NOVO NORDISK A/S) 08 AUGUST 1996. SEE ENTIRE DOCUMENT.	1-26, 32, 36- 40, 46, 50-52 27-29, 41-43, 75-76																							
X	WRIGHT, D.E. ET AL. FLUOROMETRIC ASSAY FOR ADENOSINE 3',5'- CYCLIC MONOPHOSPHATE-DEPENDENT PROTEIN KINASE AND PHOSPHOPROTEIN PHOSPHATASE ACTIVITIES. PROC. NATL. ACAD. SCI. USA, OCTOBER 1981, VOL. 78, NO. 10, PAGES 6048-6050. SEE ENTIRE DOCUMENT.	1-4, 6-9, 11-13, 15-17, 19, 20, 23, 37, 39; 40, 43, 50-52																							
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.																									
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B	earlier document published on or after the international filing date	*Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art																						
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/12410

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	MALENCIK, D.A. et al. Characterization of a Fluorescent Substrate for the Adenosine 3',5'-Cyclic Monophosphate-Dependent Protein Kinase Anal. Biochem. 1983, Vol. 132, pages 34-40, see entire document.	1-4, 6-9, 11-13, 15-17, 19, 20, 23, 37, 39, 43, 50-52
X	ZHAO, Z. Characterization of a New Substrate for Protein Kinase C: Assay by Continuous Fluorometric Monitoring and High Performance Liquid Chromatography Biochem. Biophys. Res. Commun. 15 May 1991, Vol. 176, No. 3, pages 1454-1461, see entire document.	1-4, 6-9, 11-13, 15-17, 19, 20, 23, 37, 39, 43, 50-52
A	WO 91/01305 A1 (UNIVERSITY OF WALES COLLEGE OF MEDICINE) 07 February 1991. see entire document.	1-52, 75, 76
A	SALA-NEWBY, G.B. et al. Engineering a Bioluminescent Indicator for Cyclic AMP-Dependent Protein Kinase. Biochem. J. 1991. Vol. 279, pages 727-732, see entire document.	1-52, 75, 76

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/12410

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☒ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
1-52, 75, 76
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/12410

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (6):

C12Q 1/48, 1/68; C07K 19/00; C12N 15/62, 15/11, 1/21, 15/10, 15/09

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, MEDLINE, SCISEARCH, LIFESCI, BIOTECHDS, BIOSIS, EMBASE, CAS, NTIS, WPI
search terms: proteuin kinase#, assay? or measur?, phosphorylat?, site# or domain# or region#, fusion or insert?,
fluorescent?, green fluorescent protein# or gfp

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claims 1-36, 52, 75, and 76, drawn to a fluorescent protein substrate for a protein kinase, method of making said substrate and method of use of said substrate.

Group II, claims 37-51, drawn to nucleic acids encoding a fluorescent protein substrate for a protein kinase.

Group III, claims 53-55, drawn to peptide library.

Group IV, claims 56-74, drawn to a gene library and method of use thereof.

The inventions listed as Groups I-IV do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: The proteins of Group I and DNA of Group II comprise unrelated chemical structures. The proteins of Group I and peptide library of Group III do not share a technical feature as the proteins of Group I are not required to be a part of the peptide library of Group III. The nucleic acids of Group II and gene library of Group IV do not share a technical feature as the nucleic acids of Group II are not required to be a part of the peptide library of Group IV. Accordingly, the claims are not so linked by a special technical feature within the meaning of PCT Rule 13.2 so as to form a single inventive concept.

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